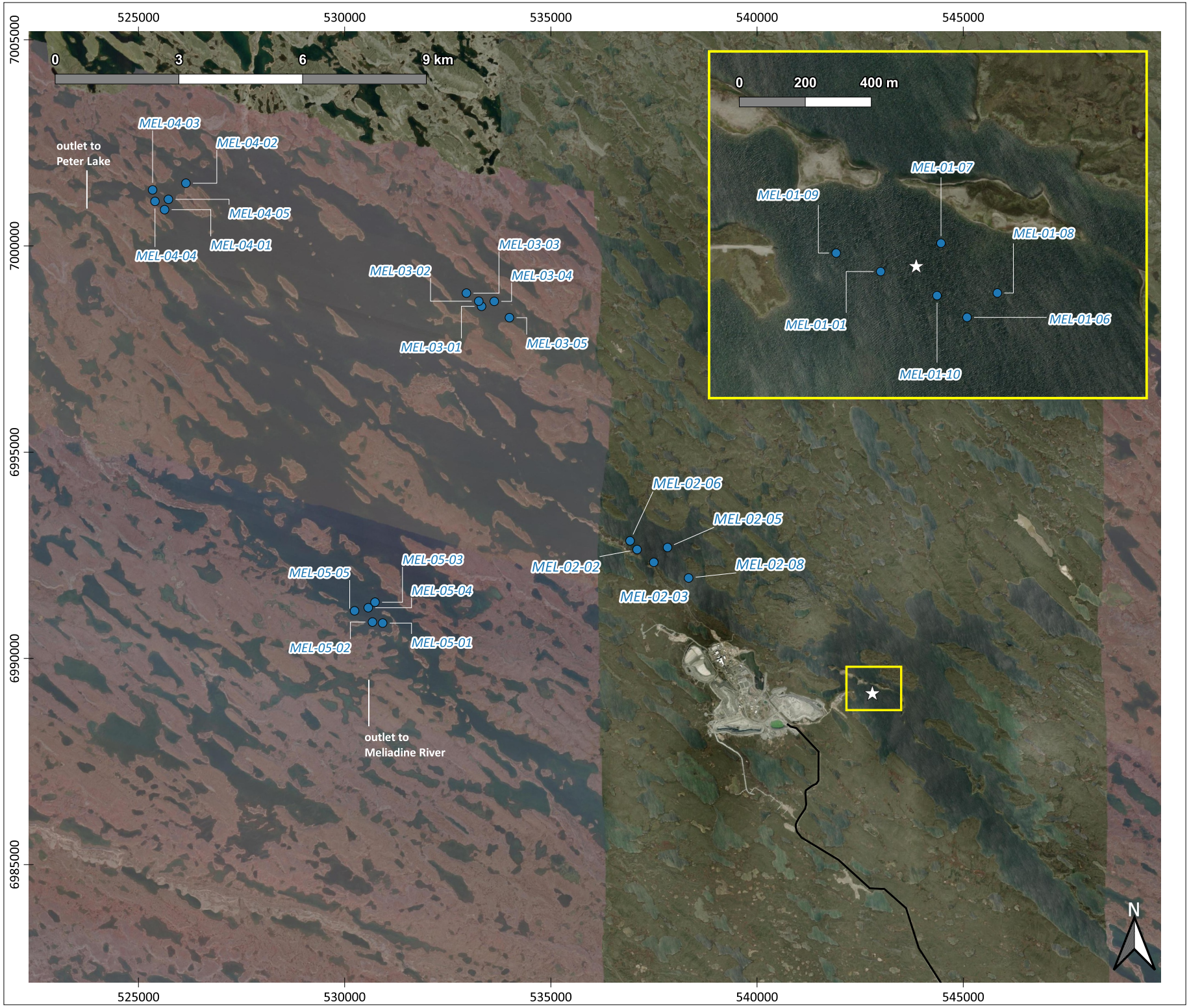

Figure 4-1
Sampling Areas and Stations for the Meliadine Lake AEMP

AEMP Design Plan (Verison 3)

AZIMUTH

Date:	February 3, 2025
Datum:	NAD 83 UTM Zone 15N
Scale:	1:119,000 ; inset =1:15,000
Software:	QGIS version 3.22.11-Białowieża
Produced by:	E. Franz

REFERENCES:
1. Basemap imagery from ESRI

Legend

- All weather access road
- ☆ Diffuser
- AEMP Sampling Stations
  - Coordinates for water/phytoplankton and sediment/benthic invertebrates are provided in Table 4-1.

Threespine Stickleback are collected from littoral areas in MEL-01, MEL-03 and MEL-04 in the vicinity of the fixed sampling stations. Lake Trout are collected from the area around the diffuser.

Table 4-1. Sampling Stations for Meliadine Lake Study (NAD 83, Zone 15V).

Area	Station ID	Water and Phytoplankton			Sediment and Benthic Invertebrates		
		Depth(m)	Easting	Northing	Depth(m)	Easting	Northing
Near-field Area water quality phytoplankton, sediment quality, benthic invertebrates	MEL-01-01	9.4	542690	6989132	9	542674	6989120
	MEL-01-06	8.8	542952	6988993	8.9	542739	6989050
	MEL-01-07	7.7	542873	6989218	8.7	542876	6989070
	MEL-01-08	7.5	543044	6989067	8.5	543064	6989183
	MEL-01-09	7.1	542555	6989188	7.9	542552	6989120
	MEL-01-10	10.5	542861	6989059	-	-	-
Mid-field Area water quality phytoplankton, sediment quality, benthic invertebrates	MEL-02-02	10.0	537093	6992642	10	537103	6992630
	MEL-02-03	9.8	537497	6992332	9.8	537497	6992327
	MEL-02-05	9.4	537831	6992692	9.4	537774	6992496
	MEL-02-06	10.2	536922	6992853	10.2	536951	6992914
	MEL-02-08	9.7	538342	6991952	9.7	538324	6991957
Reference Area 1 water quality phytoplankton, sediment quality, benthic invertebrates	MEL-03-01	9.5	533321	6998540	9.5	533492	6998645
	MEL-03-02	10.5	533253	6998664	10.5	533310	6998690
	MEL-03-03	10.5	532954	6998860	10.5	532989	6998869
	MEL-03-04	8.0	533629	6998660	8	533580	6998653
	MEL-03-05	8.1	533997	6998265	8.1	533999	6998274
Reference Area 2 water quality phytoplankton	MEL-04-01	8.3	525634	7000884	-	-	-
	MEL-04-02	9.8	526151	7001525	-	-	-
	MEL-04-03	10.7	525343	7001363	-	-	-
	MEL-04-04	8.9	525401	7001085	-	-	-
	MEL-04-05	8.5	525727	7001134	-	-	-
Reference Area 3 water quality phytoplankton, sediment quality, benthic invertebrates	MEL-05-01	9.6	530922	6990859	9.6	530716	6991054
	MEL-05-02	9.8	530675	6990883	9.8	530692	6990913
	MEL-05-03	8.6	530737	6991365	8.6	530726	6991399
	MEL-05-04	9.9	530573	6991231	9.9	530658	6991206
	MEL-05-05	10.5	530241	6991156	10.5	530305	6991196

Note:
Station locations shown above were from the 2021 AEMP. The exact UTMs may vary slightly year-to-year for the fixed monitoring stations. Sediment and benthic invertebrate sampling locations are collocated with water and phytoplankton were possible. If habitat differences are present, the stations are relocated to more suitable sampling locations.
Sediment and benthic invertebrate community sampling were discontinued at MEL-04 in 2018 based on differences in habitat in this area of Meliadine Lake (Golder, 2019).

Table 4-2. Sampling and Analysis Plan for the Meliadine Lake AEMP.

Component	Monitoring Areas	Frequency	Timing	Samples Per Event	Parameters/Endpoints
Effluent Chemistry	MEL-14	Annual	Weekly during discharge	1	Chemistry: as per MDMER and the Water Licence
Acute Toxicity	MEL-14	Annual	Monthly during discharge	1	Rainbow Trout and *Daphnia magna*
Sublethal Toxicity	MEL-14	Annual	Up to 2 times per year	1	Lemna minor growth inhibition[a]
Surface Water	MEL-01, MEL-02	Annual	March or April + July, August, September	6 for MEL-01; 5 for MEL-02	Field measurements, full suite of laboratory parameters (e.g., major ions, nutrients, metals, cyanide)
	MEL-03	Annual	July, August, September	5 per area	
	MEL-04, MEL-05	Annual	August	5 per area	
Phytoplankton	All Meliadine Areas	Annual	August	6 for MEL-01; 5 for the other areas	Phytoplankton community (biomass and density at the lowest practical level of identification) chlorophyll-a
Benthic Invertebrates and Sediment	MEL-01, MEL-02, MEL-03, and MEL-05	3-year cycle	August	5 per area	Benthic invertebrate abundance at the lowest practical level of identification Sediment chemistry (grain size, TOC, metals, nutrients)
Threespine Stickleback Population	MEL-01, MEL-03, MEL-04	3-year cycle	August	Lethal Survey: approximately 25 mature male and female fish per area[]	Age, length, weight, condition, sex, fecundity, size at age, external and internal health (including gonad and liver weights)
Lake Trout Population	MEL-01, Atulik Lake, Peter Lake	3-year cycle	August	Lethal Survey: approximately 30 fish combined for both sexes[b]	Age, length, weight, condition, sex, fecundity, size at age, external and internal health (including gonad and liver weights)
Threespine Tissue Chemistry	MEL-01, MEL-03, MEL-04	3-year cycle	August	10-20 fish in each area	Carcass (viscera removed) Metals, moisture,
Lake Trout Tissue Chemistry	MEL-01, Atulik Lake, Peter Lake	3-year cycle	August	Approximately 20-30 fish across a range of size classes	Muscle (liver and kidney archived) Metals, moisture,

Notes
[a] The most sensitive test species based on sublethal toxicity test results (2018 to 2020).
[b] Subject to refinement based on input from the Technical Advisory Panel in their review of the Cycle 3 EEM program.

Table 4-3. Summary of Fish Captured in Meliadine Lake and Potential Reference Lakes (1997 to 2013) Using Various Capture Methods

Lake	Meliadine Lake	Potential Reference Lakes				
		Atulik Lake[a]	Chickenhead Lake	Control Lake	Little Meliadine Lake	Parallel Lake
Large-bodied Fish						
Arctic Char	473	0	0	0	30	0
Arctic Grayling	199	0	12	2	83	0
Burbot	19	0	1	1	1	0
Cisco	2,503	0	0	0	27	6
Lake Trout	463	0	17	16	83	38
Lake Whitefish	0	0	0	0	0	1
Round Whitefish	114	0	0	42	91	19
Small-bodied Fish						
Ninespine Stickleback	0	0	0	38	18	0
Threespine Stickleback	6,243	0	0	0	0	0
Slimy Sculpin	4	0	0	1	7	0

Notes:
[a] From Volume 7 of the 2014 FEIS (Agnico Eagle, 2014).

4.2 Effluent Characterization and Surface Water Quality

4.2.1 Revisions in Version 3

Minor edits were made to water quality monitoring program in Version 3 of the AEMP Design Plan to address comments that were received from ECCC regarding the January 2024 version of the AEMP Design Plan. The following revisions apply to the Meliadine Lake water quality component of the AEMP:

ECCC-TC-08: Water quality screening criteria for parameters without CCME guidelines

ECCC recommended the Proponent update the water quality screening criteria in both the Water Balance and Water Quality Model and Aquatic Effects Monitoring Plan Design Plan, to include Federal Environmental Quality Guidelines (FEQGs) for cobalt, copper, strontium and vanadium. FEQGs for vanadium (2016) and cobalt (2017) were incorporated into the AEMP for water quality screening in 2019 (Azimuth 2020). FEQGs for other parameters may be added as AEMP Benchmarks in the future.

ECCC-TC-17: Parameter concentration normal ranges in Meliadine Lake

ECCC had several questions pertaining to the Normal Range assessment in Meliadine Lake, including data and methods used to calculate the updated Normal Ranges in 2020. The temporal and spatial trend assessment in **Section 4.2.5** was updated to provide clarity about the baseline/reference data and methods used to derive the Normal Ranges for Meliadine Lake.

ECCC-TC-18: Comparison between observations and FEIS predictions

The January 2024 version of the AEMP Design Plan proposed comparing current water quality results in the East Basin "against the most up-to-date water quality model predictions as that information becomes available." ECCC recommended comparing observed water quality at MEL-01 against predictions in the 2014 FEIS and against updated water quality models as they come available.

4.2.2 Objectives

The primary objectives of the water quality component of the Meliadine Lake study are as follows:

- Characterize effluent quantity and quality at MEL-14 to assess compliance with MDMER and Water Licence requirements and to support interpretation of effects in the receiving environment,
- Characterize water quality at the edge of the mixing zone and within Meliadine Lake to assess compliance with Water Licence requirements, meet MDMER requirements, and to support interpretation of effects in the receiving environment,
- Determine whether the Mine is causing changes to water quality in Meliadine Lake,
- Evaluate the accuracy of predicted changes in water quality,

- Assess whether mitigation measures are effective at reducing impacts to the aquatic environment, and
- Provide recommendations (as required) for follow-up monitoring or mitigation to lower the impact of mining-related activities on changes in water quality.

These objectives are addressed by answering the following key questions:

- Are concentrations of parameters in the effluent less than limits specified in the Water Licence?
- Has water quality in the exposure areas changed over time, relative to reference/baseline areas?
- Is water quality consistent with predictions outlined in the FEIS and are concentrations less 75 % of the applicable water quality guidelines set as AEMP Benchmarks)[3]?

4.2.3 Study Design and Schedule

Effluent Characterization

Effluent quality samples are collected according to MDMER and Water Licence requirements. Samples for effluent characterization are collected at MEL-14 located in the Effluent Water Treatment Plant (EWTP). This is the regulated monitoring station and at the last point of control before surface contact water is discharged to Meliadine Lake. The parameters and schedule for the effluent quality monitoring program are shown in **Table 4-4**. More detailed information on effluent sampling and water quality sampling for compliance and verification monitoring purposes can be found in the current Water Management Plan and Water Quality and Flow Monitoring Plan.

Meliadine Lake Receiving Water Quality

Details regarding the Meliadine Lake water sampling program are provided in **Table 4-5**. Four sampling events are conducted annually at MEL-01 and MEL-02 each year: one under ice event in March or April and three open-water events in July, August, and September. Reference Area 1 (MEL-03) is sampled in July, August, and September, while Reference Areas 2 and 3 (MEL-04 and MEL-05) are sampled only in August.

Water samples are collected at five stations in each area, except for MEL-01, where six locations are sampled around the diffuser. Three stations (MEL-01-01, MEL-01-07, and MEL-01-10) are located 100 meters from the diffuser, while another three stations (MEL-01-06, MEL-01-08, and MEL-01-09) are positioned 250 meters from the diffuser. This station configuration serves two purposes: (1) to verify

[3] The AEMP Benchmarks correspond to the lowest water quality guidelines for the protection of aquatic life and human health, or site-specific water quality objectives in the case of fluoride, arsenic, and iron. AEMP Benchmarks are listed in **Appendix B**.

that water quality meets FEIS predictions at the edge of the mixing zone and (2) to determine the spatial extent of changes in water quality.

4.2.4 Field and Lab Methods

At each water quality station, specific conductivity, dissolved oxygen (DO; concentration and percent saturation), pH, and water temperature are recorded using a water quality multi-meter. Measurements are taken near the surface and at 1-meter intervals from the surface to just above the sediment. Secchi depth is measured during open-water conditions to provide a visual assessment of water clarity.

During winter programs, ice thickness is measured at each station. Total water depth below the ice is then measured with a sounding line or an equivalent method.

Additional field-recorded information includes total water depth, station coordinates, date and time of sample collection, sample collection depth, and weather conditions.

Table 4-4. Meliadine Lake Effluent Characterization Details at the Point of Discharge (MEL-14) and Edge of Mixing Zone (MEL-13)

Location (Station ID)	Parameters[a]	Frequency
EWTP (MEL-14)	Volume (m^3)	Daily during periods of discharge
	Field effluent quality measurements	Weekly during periods of discharge
	Parameters as listed in 'Schedule I Full Suite' and 'Group 3 (MDMER and the Water Licence)	Prior to discharge and weekly during periods of discharge
	Acute toxicity testing Rainbow Trout & *Daphnia magna*	Once prior to discharge and monthly during discharge
	Lemna minor sublethal toxicity testing as per MDMER[b]	Quarterly
Receiving Environment at the Diffuser (MEL-13)	Field measurements and 'Schedule I Full Suite' and 'Group 3 (MDMER and the Water Licence)	Monthly during discharge

Notes:
[a] Detailed parameter list in **Table 4-6**.
[b] Schedule 5, Part 1, Section 6(3): After three years, sublethal testing can be conducted once per calendar quarter on test species that with the lowest inhibition concentration that produces a 25% effect or an effective concentration of 25%.

Table 4-5. Meliadine Lake Receiving Water Quality Design Plan Details

Location	Stations per area	Parameter[a]	Sampling Frequency
Near-field (MEL-01)	6	Field measurements and parameters as listed in 'Schedule I Full Suite' and 'applicable Group 3 (MDMER)' of the 2AM-MEL1631 NWB Water Licence	Annual; March/April and July, August, and September
Mid-field (MEL-02)	5	Field measurements and parameters as listed in 'Schedule I Group 2'	
Reference Area 1 (MEL-03)	5	Field measurements and parameters as listed in 'Schedule I Group 2' and 'applicable Group 3 (MDMER)' of the 2AM-MEL1631 NWB Water Licence	Annual; July, August, and September
Reference Area 2 (MEL-04)	5	Field measurements and parameters as listed in 'Schedule I Group 2' of the 2AM-MEL1631 NWB Water Licence	Annual; August
Reference Area 3 (MEL-05)	5		

Notes:
[a] Detailed parameter list in **Table 4-6.** Further details in Water Licence (2AM-MEL1631).

Water samples are collected from approximately mid-depth in the water column using a Kemmerer sampler (or equivalent) during the open-water season, and with an electric diaphragm pump with tubing during the ice-cover season. Sample bottles are provided by an accredited analytical laboratory and samples will be processed (i.e., filtered and/or preserved as required, and refrigerated) according to the instructions provided by the laboratory. Water samples requiring filtration will be filtered through a 0.45 µm syringe filter and preserved according to specifications from the lab. Water samples will be kept refrigerated before shipping and ice-packs will be added to the coolers during transport. Samples will be shipped to the analytical laboratory as soon as feasible after sample collection and processing. Quality control samples (duplicate and blanks) will be collected at randomly selected stations to represent at least 10% of all samples collected. Effluent samples will be collected for chemical analysis as per the Water Licence at the effluent water treatment plant discharge location (MEL-14).

The suite of parameters to be analyzed in the water quality samples is listed in **Table 4-6**. Water quality samples will be analyzed by an accredited laboratory at detection limits lower than applicable water quality guidelines. The corresponding information for effluent quality sampling is provided in the Water Licence and Water Quality and Flow Monitoring Plan.

Table 4-6. List of Water Quality Parameters

Group	Parameters
Field	Field pH, specific conductivity, dissolved oxygen, and temperature, Secchi depth (open-water), total depth, ice thickness (winter)
Group 2	*Conventional Parameters*: bicarbonate alkalinity, chloride, carbonate alkalinity, turbidity, conductivity, hardness, calcium, potassium, magnesium, sodium, sulphate, pH, total alkalinity, total dissolved solids (TDS; calculated [a,b]), total suspended solids (TSS), total cyanide, free cyanide, and weak acid dissociable (WAD) cyanide *Nutrients*: ammonia-nitrogen, total Kjeldahl nitrogen, nitrate-nitrogen, nitrite-nitrogen, ortho-phosphate, total phosphorus, total organic carbon, dissolved organic carbon, and reactive silica *Total and dissolved metals*: aluminum, antimony, arsenic, barium, beryllium, boron, cadmium, chromium, copper, iron, lead, lithium, manganese, mercury, molybdenum, nickel, selenium, silver, strontium, thallium, tin, titanium, uranium, vanadium, and zinc
Group 3 / MDMER	*Deleterious Substance*: pH, temperature, TSS, metals (arsenic, copper, lead, nickel, zinc), cyanide, radium-226[c], and un-ionized ammonia[d] *MDMER parameters*: conductivity, turbidity, hardness, alkalinity, chloride, nitrate, total ammonia, phosphorus, sulphate, aluminum, cadmium, chromium, cobalt, iron, manganese, mercury, molybdenum, selenium, thallium, uranium
Full Suite	Group 2, total petroleum hydrocarbons, and turbidity

Notes
[a] Standard Methods (Method 1030E, APHA 20121).
[b] TDS calculated (mg/L) = (0.6 x Total Alkalinity as CaCO3) + Sodium + Magnesium + Potassium + Calcium + Sulfate + Chloride + Nitrate + Fluoride + Silicate
[c] Sampled as part of the MDMER sampling at the Near-field area and Reference Area 1. Monitoring of radium-226 is not required if concentration in effluent is lower than 0.037 Bq/L for 10 consecutive weeks (MDMER; Schedule 5; Part 1, Section 7(d)(ii)).
[d] Un-ionized ammonia is not listed in the Water Licence, but it is included in the list of Prescribed Deleterious Substances in the MDMER.

4.2.5 Data Analysis and Interpretation

Effluent Characterization

Effluent samples are screened against the MDMER limits for deleterious substances and concentration limits in the Water Licence. The results from acute and sublethal toxicity testing on the final effluent will also be reported to meet these requirements. Standard endpoint calculations and associated parameters (e.g., LC50 and IC25 results) will be completed by the laboratory and reviewed before reporting in the AEMP.

Meliadine Lake Receiving Water Quality

The water quality assessment for Meliadine Lake includes the following elements: (1) screening against aquatic life and human health drinking water guidelines (AEMP Benchmarks), assessing temporal and spatial trends, and evaluating current water quality against predictions in the FEIS.

Water Quality Screening Assessment (AEMP Benchmarks)

AEMP Benchmarks refer to water quality guidelines for the protection of aquatic life, guidelines for the protection of human drinking water quality, or site-specific water quality objectives (SSWQO) developed for the Mine (arsenic, fluoride, iron). Water chemistry results are screened against the AEMP Benchmarks each year. To provide an added level of protection, 75 % of the AEMP Benchmark is used as an early warning 'trigger' as part of the adaptive management strategy (**Section 6**).

To simplify the screening assessment, the lowest of the freshwater aquatic life and drinking water guidelines for each parameter are adopted as the AEMP Benchmark (and corresponding trigger). Except for fluoride, arsenic, and iron, which have SSWQOs, and antimony which has a lower health-based drinking water quality guideline, the aquatic life guidelines are more conservative (i.e., lower). Therefore, if the concentration of a given parameter is below the AEMP Benchmark for aquatic life, the Benchmark for drinking water quality is also met.

AEMP Benchmarks for toxicological effects on aquatic life are adopted from the most recent guidelines published by the following sources:

- Canadian Council of Ministers of the Environment (CCME) – The freshwater aquatic life guidelines published by CCME were adopted as the AEMP Benchmarks for the protection of aquatic life unless more recent guidelines are available.
- Federal Environmental Quality Guidelines (FEQG) – FEQGs are being developed for parameters where the CCME guidelines for the substance have not yet been developed or are not reasonably expected to be updated in the near future. FEQGs are similar to CCME WQGs in that they are based on toxicological effects data using the same methods of derivation, where adequate data exists. Parameters with more recent FEQG include vanadium (2016), cobalt (2017), lead (2020), strontium (2020), copper (2021), and aluminum (2022).
- Guidelines published by the British Columbia Ministry of Environment and Climate Change Strategy (BC ENV) for parameters not covered under either CCME or FEQGs (e.g., sulphate).
- Guidelines from other jurisdictions (e.g., TDS guideline for Alaska of 500 mg/L [ADEC 2012]).
- Canadian drinking water quality guidelines (Health Canada, 2020).

Temporal and Spatial Trends

Temporal and spatial trends are evaluated by comparing water quality results from the open water period to the Normal Range and visually examining the data. Appropriate statistical methods are incorporated into the assessment to support the discussion, as needed. Normal range refers to the natural water quality conditions in Meliadine Lake. Provisional Normal Ranges for Meliadine Lake were calculated in 2018 using the available baseline and reference data throughout Meliadine Lake. Not enough sampling was completed during the baseline period to calculate Normal Ranges for different

basins of Meliadine Lake. Therefore, the baseline and reference dataset for the entire lake was pooled to derive one set of Normal Range values. The authors expressly stated that the Normal Ranges would be updated to include new reference area data (see page iii in the Executive Summary of the 2018 AEMP and Cycle 1 EEM [Golder, 2019]). The Normal Ranges were updated for the 2020 AEMP to include reference area samples from MEL-03, MEL-04, and MEL-05 in 2019 and 2020. For most parameters, the updated Normal Ranges equal to the upper 90th percentile concentration measured in samples collected throughout Meliadine Lake during the open water period from 1995 to 2013 and samples collected from the reference areas to the end of 2020. For parameters that were routinely measured below the analytical detection limit, the Normal Range was set equal to the analytical detection limit. The methods used to calculate the Normal Range for each parameter are presented in the 2019 AEMP report (Azimuth, 2020).

Spatial and temporal trends are evaluated primarily using data collected during open water season for three reasons: (1) reference areas are not sampled during the winter, (2), conditions under ice at the reference areas were not characterized during the baseline period, (3) effluent is only discharged during the open water period.

A generalized workflow was developed to short-list the number of parameters that are carried forward for closer examination.

- Parameters with fewer than 50% detected concentrations are excluded from the spatial and temporal trend assessment. Monthly water quality results are examined to verify that the frequency of non-detects is consistent in each month.
- Normal Range Assessment – Parameters that exceed the Normal Range in any of the samples collected from MEL-01 or MEL-02 in the current year are added to a “long list”. Sample-by-sample screening is a coarse tool for assessing changes in water quality because in any given event there may be results that naturally exceed the normal range. Parameters measured in water from the MEL-01 and MEL-02 are considered outside the Normal Range if the mean or median concentration from the open water period exceeds the Normal Range. The mean calculation can be affected by outliers (high or low concentrations) that do not influence the median. Therefore, the median concentration serves as a “check” for potential outliers in the Normal Range assessment.

- Statistical Analyses– Water quality parameters that exceeded the Normal Range were carried forward for quantitative analysis of year-over-year differences within MEL-01, MEL-02, and MEL-03 using analysis of variance (ANOVA) and Tukey post-hoc pairwise comparisons (significant difference at $\alpha = 0.05$). This assessment focuses on data from MEL-01, MEL 02, and MEL 03 because these three areas are sampled monthly during the open water season. The magnitude of year-to-year changes in water quality parameter within each area is calculated using the model estimates for each water quality parameter.

The Normal Range assessment and statistical analysis help to differentiate parameters that are elevated compared to baseline/reference but stable in recent years versus those parameters that show consistent year-over-year increases related to mining activities, wider regional patterns of change, or a combination of factors.

Comparison to Predictions in the FEIS

An important aspect of the water quality assessment for Meliadine Lake is determining if the pattern, timing, and magnitude of changes in water quality generally align with the predicted changes based on the approved design plan for the Mine. Predicted future changes in water quality also provide a point of comparison with which to evaluate how effectively the Mine is managing water quality on site. Parameters that are increasing over time are compared against the prediction presented in the 2014 FEIS:

Water quality in the east basin of Meliadine Lake is predicted to change relative to baseline conditions, but aquatic life and health-based guidelines would be met at 100 m from the diffuser.

The narrative statement of "water quality meeting guidelines at the edge of the mixing zone" was based on modelling of effluent mixing and dilution estimates completed as part of the FEIS in 2014. Predicted concentrations were developed for several parameters at the edge of the mixing zone, as well as for TDS, chloride, and sodium beyond the mixing zone in the east basin. The model was based on the extent of the approved mine plan in the 2014 FEIS, conservative assumptions regarding effluent quality, and the preliminary diffuser design. The *far-field*[4] effluent mixing model in Volume 7 of the FEIS predicted TDS, chloride, and sodium would increase gradually over time in the East Basin to maximum concentrations of 176 mg/L for TDS, 66 mg/L for chloride, and 19 mg/L for sodium in the last year of operations. Water quality data collected from MEL-01 stations will be compared to water quality predictions in the 2014 FEIS and updated water quality model results as those data becomes available.

[4] Far-field in this case means the broader east basin. This is not to be confused with the reference areas in Meliadine Lake.

4.2.6 Quality Assurance and Quality Control

Quality assurance and quality control procedures determine data integrity and are relevant to sample collection through to data analysis and reporting. Quality assurance (QA) encompasses management and technical practices designed at the outset to confirm that the data generated are of consistent, acceptable quality. Quality control (QC) is an aspect of QA and includes the procedures used to measure and evaluate data quality, and the corrective actions to be taken when data quality objectives (DQOs) are not met.

A summary of QA/QC procedures for assess data quality for the water chemistry monitoring program are presented below. These procedures are used to confirm that the water quality data are representative of known quality, properly documented, and scientifically defensible.

Field Collection

Samples will be collected by qualified field staff who have received appropriate training. Fieldwork will be completed according to approved specific work instructions and established technical procedures. Specific work instructions are standardized forms that describe exact sampling locations and provide specific sampling instructions, equipment needs and calibration requirements, sample labelling protocols, shipping protocols, and laboratory contacts.

Careful documentation and handling of samples and data is a key component of QA/QC for the water quality field program. Sample containers are labeled with the sample ID, the date, and project identification. They are kept or stored according to laboratory handling instructions as necessary. Field data are recorded on data sheets and entered in Agnico Eagle's EQuIS database. Field data are sent to Azimuth at the end of each sampling event and used to validate data entry in EQuIS.

Chain-of-custody forms are included in each shipment. Electronic copies are emailed to the account manager when samples leave the Site. Samples are typically shipped within one week of collection, typically on Monday, Tuesday, or Wednesday to avoid having samples in transit over a weekend.

Laboratory QC

ALS Environmental is a CALA[5] certified laboratory with a rigorous QA/QC system that includes:

- Setting holding times according to test methods and any exceedances are flagged.
- Determining detection limits (DL), which is the minimum concentration of an analyte detectable by a test method in a medium and values below this limit are reported as less than DL.
- Including several QA/QC samples in their standard analytical procedures:

[5] Canadian Association for Laboratory Accreditation

  - Matrix spikes are a quality assurance measure used to determine the resolution of a test method to detect an analyte in a specific medium (matrix) and assess matrix interferences.
  - Matrix blanks are analyzed to assess background contamination that exists in the analytical system that could lead to elevated concentrations or false positive data. These samples are comprised of analyte-free water.
  - Laboratory control samples are comprised of a mixture of analyte-free water to which known amounts of the method analytes are added. They are essentially an internal version of certified reference material.
  - Certified/standard reference materials are commercially-made with pre-determined analyte concentrations and are sampled systematically to ensure accuracy.
- Analysis of laboratory replicate samples to determine variability in reported analyte concentrations.
- Verifying reports by repeat analysis of a sample if the original result is unexpected (e.g., detecting a parameter in blank samples and deviations from historical results). Repeat analysis may be requested by the client or consulting team.

Data Quality Objectives (DQOs) are numerically definable measures of analytical precision and completeness. Analytical precision is a measure of the variability associated with duplicate analyses of the same sample in the laboratory. Laboratory duplicate results are assessed using the relative percent difference (RPD) between measurements. The equation used to calculate the RPD is:

$$RPD = \frac{(A - B)}{(\frac{A + B}{2})} x\ 100$$

where: A = analytical result; B = duplicate result.

RPD values may be either positive or negative, and ideally should provide a mix of the two, clustered around zero. Consistently positive or negative values may indicate a bias. Large variations in RPD values are often observed between duplicate samples when the concentrations of analytes are very low and approaching the detection limit; and therefore, a difference (DIFF) metric is often relied upon in these cases. The DIFF metric is defined as the absolute difference between a sample result and the sample duplicate result for each analyte.

$$DIFF = ABS\ [A - B]$$

where: A = analytical result; B = duplicate result; ABS = Absolute value (i.e., positive).

The chemistry laboratory DQOs for this project are:

- Analytical precision targets set by the lab are parameter-specific but typically are approximately 20% RPD or a difference (DIFF) between the laboratory replicates of greater than 2-times the DL (or in some cases 3-times the DL); meeting either metric is acceptable. If the RPD or DIFF metrics are not met, the result is flagged.
- Other QA/QC metrics flagged by the laboratory are evaluated to determine any implications on chemistry results. These include: laboratory holding time, laboratory control sample, matrix spike, method blank, certified/standard reference materials, detection limit, and reported result verified by repeat analysis.

Field QC

The standard QA procedures included thoroughly rinsing sampling equipment between stations to prevent cross-contamination. Field QC procedures include collecting and analyzing field duplicates, and three types of *blank* samples: travel blanks, field blanks (de-ionized water), and equipment blanks.

Field Duplicates

An independent collection of water samples at the same time and location as the original, as a measure of consistency in sampling methodology and heterogeneity of chemical parameters at discrete locations. One field duplicate is collected for every 10 samples (approximately 10% frequency).

The DQOs for field duplicates were 1.5-times the laboratory RPDs or the DIFF between field duplicate results of less than 3-times the DL (i.e., 1.5x the difference objective for laboratory duplicates). This approach has been adopted for both water chemistry and sediment chemistry since 2019. The adjustment of field DQOs above laboratory RPD levels accounts for the fact that field duplicates are inherently more variable compared to laboratory duplicates partly because field duplicate samples are collected from a large sample volume as opposed to a small well-mixed sample volume (i.e., the single sample container in the laboratory). The Canadian Council of Ministers of the Environment (CCME) states that acceptance limits for field-based QC are broader than laboratory QC and are typically 1.5 to 2 times the laboratory QC limits (CCME, 2016).

Blanks

Three types of “blanks” are collected as part of water quality QC assessment according to best practices and guidance published by BC Ministry of Environment (2013) and CCME (2011).

- Travel Blanks – Travel blanks, or trip blanks, consist of de-ionized (DI) water provided in sampling bottles by ALS and receive the same treatment as field samples during shipment, handling, storage, and laboratory analysis. Trip blanks are meant to detect any widespread contamination resulting from the container (including caps) and preservative during transport and storage. Travel blanks should (1) be included in sample container shipments, (2) come directly from the analytical laboratory and (3) be stored in a cool place (e.g., refrigerator).
- Field Blank (*aka deionized water blank [DI blank])* – Laboratory-supplied deionized water is poured directly into the sample bottles. Field blanks are used to detect potential contamination caused by from bottles, collection methods, the atmosphere, and preservatives. The field blank mimics the water sample except the deionized water does not come in contact with the sampling device (pump and tubing in the winter and Kemmerer during the open water season).
- Equipment Blanks – At the beginning or end of a field sampling episode, after routine rinsing of the pump and tubing or Kemmerer, distilled water is run through the equipment and placed in sampling bottles for analysis of a wide suite of parameters (e.g., metals, nutrients, and major ions). This sample tests for possible cross-contamination of samples from the water sampling equipment.

Blank sample collection, particularly equipment blank samples, required careful planning, attention to detail, focus on the importance of cleanliness, and generally provided a good opportunity to refine sample collection skills. Blank samples are collected once per sample event and submitted blind to the laboratory to ensure they were treated the same as field-collected samples during analysis.

Blanks are examined for detectable concentrations of any of the parameters measured. Ideally, no parameter in either blank should exceed laboratory DLs. If a parameter in either blank is detectable, the corresponding field sample results are assessed for their reliability in the water chemistry dataset. The approach utilized is a "5 x blank censoring approach", relying primarily on the EB[6] for each event, and using the following rating system for detected analytes in blanks:

- Unreliable – When the concentration in a field sample is within 5-times the concentration in the EB blank, and the field result is elevated relative to historical data for the station, results are deemed unreliable (potentially impacted by cross-contamination). These data are excluded from data analysis and interpretation.

[6] If a parameter was detected in both the EB blank and DI blank, then the detected concentration in the DI blank was subtracted from the EB blank, before comparing EB blank concentrations to field sample results.

- Cautionary – When the concentration in a field sample is less than 5-times higher than the detected analyte concentration in the EB blank, but the field result appears consistent with historical data for this lake/basin, results are flagged as cautionary. Results are considered within natural variability and are retained for data interpretation.
- Reliable – When the concentration in a field sample is more than 5-times higher than the detected analyte concentration in the EB blank or is less than the DL, the field result is considered reliable. These data are retained for data interpretation with no denotation in the tables and figures. If only the DI has a detected parameter (not EB), results are considered reliable. Reliable flags are documented in the QA/QC screening table.

The approach to evaluating blanks has been standardized to the extent possible, but ultimately best professional judgement is used to determine which data get excluded from analysis.

4.3 Phytoplankton

Phytoplankton and zooplankton monitoring were included as targeted studies in Version 1 of the AEMP Design Plan. The targeted plankton study included sampling and analysis of depth-integrated nutrients, chlorophyll a, phytoplankton, and zooplankton over three years in Meliadine Lake (2015, 2016, and 2017) and two years in the Peninsula Lakes[7] (2015 and 2016) (Golder, 2018). Phytoplankton studies have provided meaningful insight into the structure and function of the phytoplankton community in Meliadine Lake as the Mine transitioned from the pre-construction phase (2015) to operations. Furthermore, as the only biological monitoring program conducted annually under the AEMP, the phytoplankton study provides important information on the health of the aquatic environment in Meliadine Lake in years when fish and benthic invertebrate studies aren't completed as part of the 3-year AEMP and EEM cycle (2018, 2021, 2024, etc.).

4.3.1 Revisions in Version 3

The only notable update to the phytoplankton study for Version 3 of the AEMP Design Plan is a reduction in the intensity of chlorophyll-a sampling (composite) during the August sampling event. Instead of collecting triplicate composite samples at each station, one composite sample will be collected to pair with the phytoplankton taxonomy results. Variability in the chlorophyll-a concentrations within each station is low, which justifies collecting one sample per station.

[7] Chlorophyll a was also sampled at the peninsula lakes in 2017.

4.3.2 Objectives

The primary objective of phytoplankton monitoring program is to determine whether treated Mine effluent has potential short or long-term effects on phytoplankton communities. Specific monitoring objectives are as follows:

- Compare phytoplankton community metrics (i.e., chlorophyll a, phytoplankton abundance, biomass, and composition of major taxonomic groups) among areas and between years,
- Provide data to inform adaptive management intended to reduce or eliminate Mine-related effects to phytoplankton communities in Meliadine Lake, and
- Provide recommendations for future changes to the AEMP.

4.3.3 Study Design and Schedule

Phytoplankton monitoring is conducted in August at the water quality sampling locations at the five study areas in Meliadine Lake. August was selected as the most appropriate month due to lower variability in phytoplankton monitoring endpoints compared to other sampling events (Golder, 2018). Depth-integrated water samples will also be collected at these locations for analysis of chlorophyll-a.

4.3.4 Field and Lab Methods

At each sampling station, Secchi depth, total water depth, and limnology profiles will be measured prior to the collection of the plankton samples (see **Section 4.2.4** for details). After these measurements are taken, a depth integrated sample for phytoplankton and chlorophyll-a will be collected from the euphotic zone. The euphotic zone is defined as the extent of the water column that is exposed to sufficient sunlight for photosynthesis to occur (typically to a depth in the water column where 1% of the surface irradiance is measured). In the field, the euphotic zone will be calculated as two times the Secchi depth (Koenings and Edmundson 1991; Alberta Environment [AENV, 2006]). Once the euphotic zone depth is determined, a Kemmerer sampler (or equivalent) will be used to collect discrete water samples starting at the surface, and continuing every 2 m through the extent of the euphotic zone. If the total water column depth is more than 10 m, sampling would continue every 2 m through the extent of the euphotic zone. If the total water depth is less than two times the Secchi depth, then a water sample will be collected every 2 m from the surface to 2 m above the lake-bed.

Equal volumes of water from each discrete depth will be combined into a large, clean bucket to create a composite, depth-integrated sample. From this composite sample, a single subsample will be collected for phytoplankton community analysis (i.e., enumeration and identification), and triplicate subsamples for chlorophyll-a analysis.

The phytoplankton samples are collected in 50 mL plastic vials and preserved with 3-4 drops of acidified Lugol's solution. Samples are stored in the dark and shipped to Plankton R Us, Winnipeg, Manitoba, for taxonomic identification to the lowest taxonomic level, and abundance and biomass estimates.

The subsamples for chlorophyll-a are filtered at the lab on to 47 mm glass fibre type C filters with a nominal pore size of 1.2 µm. The filters are provided by the lab. A sufficient volume of water must be filtered to discolour the filter, approximately 500 mL or more per filter. Once the filtering is complete, the filter will be taken off the tower, folded in half and put into a pre-labelled Petri dish. The volume filtered will be recorded on the data sheet as well as the sample label. Samples are wrapped in aluminum foil, to prevent light penetration, and frozen. Chlorophyll-a analysis is done at the Biogeochemical Analytical Service Laboratory at the University of Alberta, Edmonton, Alberta using spectrophotometric analysis.

4.3.5 Data Analysis and Interpretation

Phytoplankton effects endpoints (i.e., density, biomass, and community composition) are evaluated, using both statistical (quantitative) and visual (qualitative) methods, to determine if mining activities have contributed to changes to the phytoplankton community.

Temporal and Spatial Trends

Time series plots organized by sampling area were used to highlight spatial and temporal patterns in nutrients, chlorophyll-a, and phytoplankton metrics. Statistical analyses may be used to evaluate subtle differences in phytoplankton community structure between the Near-field area, Mid-field area, and the three within-lake reference areas. Phytoplankton populations grow and shrink seasonally, meaning species richness, biomass, and density are expected to vary annually, in response to regional climate patterns, and spatially in response to basin-specific factors such as morphology, timing of ice off, and nutrient status. A fundamental premise of the temporal and spatial trend assessment is the phytoplankton community in the various areas of Meliadine Lake in August will vary from year-to-year, but the near-field, mid-field, and reference area communities should follow the same pattern of change each year. If, however, the phytoplankton community in near-field and/or mid-field areas diverge from previous years and from the reference areas, it may indicate water quality is influencing the structure of the community.

Community Structure

Differences in the phytoplankton community among areas and over time are determined using non-metric multidimensional scaling (nMDS). nMDS is an ordination method that takes multidimensional taxonomic data (e.g., biomass for each taxon by station-year combination) and collapses the information into two or three dimensions that capture major patterns of variation in the underlying data. Azimuth

follows a nMDS approach based on the reference condition approach (RCA) outlined in Environment Canada (2012). The fundamental premise of RCA is that a suitably large set of baseline and/or reference data can be used to characterize unimpaired conditions in terms of a variety of biological attributes. Patterns in reference area phytoplankton community structure are examined first, to determine the range of reference conditions. Patterns in community structure at the exposure areas are explored in the context of the results for the reference areas.

Starting in 2022, analysis was performed on using the biomass data for all commonly observed individual taxa. Statistical analyses for nMDS are completed in R using the statistical package 'vegan' (version 2.5-6) according to the following workflow:

- Step 1: Biomass data are compiled for all individual samples collected since August 2013. To limit the influence of rarely observed taxa, individual taxa that accounted for less than 2% of any individual sample are excluded from the analysis. Raw biomass values are log(x+1) transformed to reduce the influence of dominant taxa. This data set is then turned into a Bray-Curtis distance matrix.
- Step 2: The nMDS is run on the Bray-Curtis matrix; Shepard plots and stress values are used to optimize results. Stress, in the context of nMDS, refers to how distorted the representation of the data are in two or three dimensions compared to the original multi-dimensionality of the data. Lower stress means a better fit of the data in the reduced dimensionality. Multiple iterations of the analysis are completed to determine which position (or ordination) of points in two or three dimensions produces the lowest stress value. The guidelines outlined in Clarke (1993) are commonly used to evaluate stress values as follows: <0.05 = excellent, <0.10 = good, <0.20 = usable, >0.20 = not acceptable. Stress of nMDS ordinations tends to increase with increasing sample size and decrease with an increasing number of dimensions, independent of the structure of the underlying data (Dexter et al., 2018). Given the large number of phytoplankton samples collected over the course of monitoring at Meliadine Lake, it is expected that stress of a suitable nMDS may exceed the threshold of 0.20. Therefore, stress is considered alongside other factors such as ease of interpretation when evaluating the potential nMDS ordinations.
- Step 3: The nMDS results are visualized by first plotting 90th, 95th and 99th percentile probability ellipses using the reference data only. The next step involves adding nMDS scores for MEL-01 and MEL-02 for each year. The 90th, 95th and 99th percentile probability ellipses provide a concise way of visualizing whether the phytoplankton community at the exposure areas are within the range of baseline/reference conditions for Meliadine Lake.

In the future, other statistical approaches may be implemented on a case-by-case basis to supplement the RCA analyses if the underlying data supports a more detailed investigation of spatial and temporal trends.

Trophic Status

Trophic status is a means of classifying estimated productivity of a lake based on concentrations of key nutrients and chlorophyll-a, and on water transparency. The three main categories of productivity are:

- Oligotrophic (low nutrients, low productivity),
- Mesotrophic (intermediate productivity), and
- Eutrophic (high nutrients, high productivity).

Three parameters are used in the classification of trophic status: total phosphorus, chlorophyll-a, and water transparency. Phosphorus is the primary nutrient used in trophic status indexes because it often limits primary productivity in freshwater systems. Chlorophyll-a is the primary pigment used for photosynthesis in phytoplankton and is used as a surrogate measure of primary production. Water transparency, measured with a Secchi disk, is also used as a coarse indicator of phytoplankton biomass.

Three trophic status indices are included in the assessment:

- Vollenweider (1968) – A general classification scheme based on ranges of TP, chlorophyll-a and Secchi depth (**Table 4-7**).
- CCME (2004) – A total phosphorus-specific scheme using trigger ranges (**Table 4-8**).
- Carlson (1977) – Independent index scores for TP, chlorophyll-a and Secchi depth (**Table 4-9**), calculated as follows:

$$TSI_{TP} = 10\,(6 - \left[\frac{\ln\,(^{48}/_{TP})}{\ln 2}\right])$$

$$TSI_{Chl} = 10\,(6 - \left[\frac{2.04 - 0.68(\ln Chl)}{\ln 2}\right])$$

$$TSI_{Secchi} = 10\,(6 - \left[\frac{\ln\,\text{Secchi}}{\ln 2}\right])$$

Table 4-7. Trophic classification for lakes based on ranges of total phosphorus, chlorophyll-a and Secchi depth (Vollenweider, 1968).

Trophic Status	Total Phosphorus (mg/L)		Chlorophyll-a (µg/L)		Secchi Depth (m)	
	Mean	Range	Mean	Range	Mean	Range
Oligotrophic	0.008	0.003 to 0.018	1.7	0.3 to 4.5	9.9	5.4 to 28.3
Mesotrophic	0.027	0.011 to 0.096	4.7	3.0 to 11.0	4.2	1.5 to 8.1
Eutrophic	0.084	0.016 to 0.386	14.3	3.0 to 78.0	2.5	0.8 to 7.0

Table 4-8. Trophic classification for lakes based on total phosphorus trigger ranges (CCME, 2004).

Trophic Status	Total Phosphorus (mg/L)
Ultra-oligotrophic (very nutrient-poor)	<0.004
Oligotrophic (nutrient-poor)	0.004 to 0.010
Mesotrophic (containing a moderate level of nutrients)	0.010 to 0.020
Meso-eutrophic (containing moderate to high levels of nutrients)	0.020 to 0.035
Eutrophic (nutrient-rich)	0.035 to 0.100
Hyper-eutrophic (very nutrient-rich)	>0.100

Table 4-9. Trophic status index and general trophic classifications for lakes (Carlson, 1977).

Trophic State Index	Total Phosphorus (mg/L)	Chlorophyll-a (µg/L)	Secchi Depth (m)	General Trophic Classification
<30 to 40	0 to 0.012	0 to 2.6	>8.0 to 4	Oligotrophic
40 to 50	0.012 to 0.024	2.6 to 20	4 to 2	Mesotrophic
50 to 70	0.024 to 0.096	20 to 56	2 to 0.5	Eutrophic
70 to 100+	0.096 to 0.38+	56 to 155+	0.5 to <0.25	Hyper-eutrophic

4.3.6 Quality Assurance and Quality Control

The QA/QC procedures will be applied during all aspects of the plankton component to verify that the data collected are of acceptable quality. Data entered electronically will be reviewed for data entry errors and appropriate corrections will be made.

Field duplicates are collected for phytoplankton to assess sampling variability and sample homogeneity. A RPD of 50% for density and biomass concentrations is considered acceptable.

As a measure of laboratory QA/QC on the enumeration method, replicate counts are performed on 10% of the samples. Replicate samples are chosen at random and processed at different times from the original analysis to reduce biases. The laboratory replicate is a new aliquot (10 mL) from the sample jar and is counted from the start in the same manner as the original aliquot (10 mL) taken from the jar.

The data will be reviewed for unusually high or low values (i.e., greater or less than 10 times typical lake values), which would suggest erroneous results. Unusually high or low results will be validated on a case-by-case basis. All invalidated data will be retained in the appendix tables, but a flag will be appended to the data indicating that the sample was considered unreliable or the results were designated as not correct due to an internal review of the data.

4.4 Benthic Invertebrates

4.4.1 Revisions in Version 3

No changes were made to the benthic invertebrate study design in Version 3 of the AEMP Design Plan.

A benthic invertebrate community study was not required for the Cycle 3 EEM program because there were no statistically significant differences for any of the effect indicators (i.e., density, richness, evenness, and Bray-Curtis dissimilarity index) in either Cycle 1 (Golder, 2019) or Cycle 2 (Azimuth and Portt, 2022).

4.4.2 Objectives

The objectives of the benthic invertebrate community monitoring program are:

- Compare benthic invertebrate communities in Near-field and Mid-field areas within Meliadine Lake relative to within-lake reference areas, based on benthic invertebrate effect endpoints to identify Project-related effects.
- Verify predictions made in the FEIS and other submissions to the NWB, as applicable, relating to benthic invertebrate communities.
- Meet the requirements of the MDMER[8].
- Recommend any necessary and appropriate changes to the benthic invertebrate community component of the AEMP for future years.
- Monitor the effectiveness of proposed mitigation.
- Provide data to inform adaptive management intended to reduce or eliminate Mine-related effects to benthic invertebrate communities in Meliadine Lake.

4.4.3 Study Design and Schedule

The benthic invertebrate community study is conducted every three years in mid-to-late August when the benthic invertebrate communities are the most diverse and stable (prior to freeze up). The benthic invertebrate study includes two exposure areas (MEL-01 and MEL-02) and two reference areas (MEL-03 and MEL-05) (**Figure 4-1**). MEL-04 was dropped as a reference area for the benthic invertebrate community study in 2018 because the sediment characteristics were deemed too dissimilar compared to the exposure areas (Golder, 2019).

[8] Benthic invertebrate community sampling was not required under MDMER in 2024 because there were no confirmed effects to the benthic invertebrate community endpoints in the previous two cycles: Cycle 1 (2018) and Cycle 2 (2021).

4.4.4 Field and Lab Methods

Sampling Methods

Benthic invertebrate samples are collected within a water depth range of approximately 7 to 10 m in areas with similar sediment composition. Sediment samples are collected using a Petite Ponar (15.24 × 15.24 cm bottom sampling area of 0.0232 m²). Five replicate samples are collected in each area (e.g., MEL-01) and each sample is a composite of five individual grabs[9]. The contents of each composite grab are sieved through a 500 µm mesh screen. Material retained in the mesh is placed into a pre-labelled container and preserved in 10% neutral buffered formalin. An internal waterproof label is added to each sample container.

Sediment grab samples are also collected for chemistry (e.g., metals, nutrients, and carbon content) and particle size distribution as described in **Section 4.5.4**. The following supporting data will be collected at each benthic invertebrate sampling station:

- Station location information (e.g., coordinates, water depth, weather observations),
- Habitat description (e.g., water clarity and colour),
- Notes on the sediment substrate (e.g., colour, texture, moisture content, odour, macrophytes),
- Benthic sample-related information (grab type, mesh size, sampler fullness, preservative), and
- Photographs of the sampling areas and representative samples as necessary.

Taxonomic Analysis

Preserved benthic invertebrate samples are sent to qualified taxonomist for processing, enumeration, and identification to the lowest taxonomic level (typically genus), using current literature and nomenclature. Organisms that cannot be identified to the desired taxonomic level (e.g., immature, or damaged specimens) are reported as a separate category at the lowest level of taxonomic resolution possible.

4.4.5 Data Analysis and Interpretation

General Approach

Benthic invertebrate effect endpoints (i.e., metrics such as invertebrate density, densities of dominant invertebrates, taxonomic richness, evenness, and similarity to reference communities) will be evaluated,

[9] Pooling of subsamples in the field to form a single composite sample for taxonomic analysis from a station is commonly done to reduce analytical cost, without an effect on study results. Analysis of data collected during EEM and AEMP surveys is based on station as the unit of replication and does not require data for separate subsamples. Analyses of separate subsamples is useful to initially evaluate within-station variation, but once the number of subsamples required is determined, collection of subsample data is no longer necessary.

using both statistical (quantitative) and visual (qualitative) methods, to determine whether changes in the benthic invertebrate community have occurred. Appropriate statistical analyses will be conducted to evaluate potential differences in benthic community structure between the Near-field area, Mid-field area, and the two within-lake Reference areas.

If changes in the benthic invertebrate community are observed, the results will be further evaluated to determine whether the changes in the benthic community are within FEIS predictions and are potentially mine-related. The magnitude and direction of change in the benthic invertebrate communities will be considered, as well results from multiple evaluation methods, and results from other monitoring components such as water and sediment quality.

Data Management

Raw invertebrate abundance data will be received from the taxonomist in electronic format. The taxonomists flag certain taxa for removal, including non-benthic organisms (e.g., Cladocera, Copepoda), meiofauna that are not reliably enumerated using 500 µm mesh sampling gear (e.g., Nematoda and Harpacticoida; Environment Canada 2012, 2014), and terrestrial invertebrates. Consistent with a recommendation by Environment Canada (2014) and the subsequent approach taken by Golder (2019), Ostracoda will also be excluded from the dataset prior to analysis because these invertebrates can be found in patches of extremely high numbers and can there therefore bias sample densities, thus affecting the benthic community analysis.

Descriptive statistics will be calculated for the above metrics, including the arithmetic mean, median, minimum, maximum, standard deviation, and standard error. Benthic community variables will be presented graphically for each sampling area to allow visual evaluation of spatial and temporal patterns. Community composition will be further represented by relative abundances (i.e., as percentage of total density) of major taxonomic groups. Changes in benthic invertebrate community composition over time at the major group level will be assessed by plotting mean relative densities of major taxa by sampling area, as stacked bar graphs.

Benthic Invertebrate Effect Endpoints

Benthic community metrics for the AEMP and EEM studies are presented in **Table 4-10**.

Total density (N/m^2) and taxa richness are determined at the lowest practical level of identification. Density and richness at the level of major taxa group (MTG; Class or Order). The five MTG are Diptera (e.g., chironomids), Oligochaeta, Amphipoda, Bivalvia (clams), and Gastropoda (snails). Species that make up a minor component of the benthic invertebrate community are classified as “Other” for the purpose of calculating summary statistics and plotting. Mayflies (Ephemeroptera) and caddisflies (Trichoptera) are excluded from the dataset to stay consistent with the approach outlined in the 2018

AEMP (Golder, 2019). These taxa are typically found in streams and rivers and are not commonly found in depositional areas in lakes.

Simpson's Diversity (1-D) considers both the abundance and taxonomic richness of the community. Values in this index range from 0 to 1 with 0 representing no diversity and 1 representing infinite diversity. D is calculated according to this formula:

$$1 - \mathrm{D} = \sum_{i-1}^{s} (p_i)^2$$

Where:

D = Simpson's Diversity

p_i = the proportion of the ith taxon at the station,

S = the total number of taxa at the station (i.e., taxa richness)

Simpson's Evenness is another way of measuring the diversity of the community that takes into consideration how the total abundance is distributed among the various taxa groups. Values range from 0 to 1, with 1 representing a community with completely equal distribution of the number of individuals among the taxa. Evenness is calculated using the density data set as follows:

$$\mathrm{E} = \frac{1}{D}\ x\ \frac{1}{S}$$

Where:

E = Simpson's Evenness

D = Simpson's Diversity (see above)

S = the total number of taxa at the station (i.e., taxa richness)

The Bray-Curtis dissimilarity co-efficient is a distance measurement that reaches a maximum value of "1" for two samples that are entirely different and a minimum of "0" for two samples that possess identical descriptors (Bray and Curtis 1957). Bray-Curtis is calculated according to this formula:

$$\mathrm{BC} = \frac{\sum_{i=1}^{n} |y_{i1} - y_{i2}|}{\sum_{i=1}^{n} (y_{i1} + y_{i2})}$$

Where:

BC = Bray-Curtis distance between sites 1 and 2,

Y_{i1} = count for taxon i at site 1,

Y_{i2} = count for taxon i at site 2, and

n = the total number of taxa at the two sites.

Table 4-10: Summary of Benthic Invertebrate Community Endpoints for the EEM and AEMP.

Variable	EEM (Family Level)[a]	AEMP (Lowest Level)
Total invertebrate density (number of organisms/m^2)	Effect Endpoint (MDMER-required)	AEMP Variable
Total taxonomic richness (number of taxa per station)	Effect Endpoint (MDMER-required)	AEMP Variable
Simpson's diversity index	Supporting Endpoint	AEMP Variable
Simpson's evenness index	Effect Endpoint (MDMER-required)	AEMP Variable
Bray-Curtis Index	Supporting Endpoint	AEMP Variable
Presence/absence by each taxon	Supporting Endpoint	Supporting Endpoint
Community composition as percentages of major taxonomic groups	Supporting Endpoint	AEMP Variable
Densities of dominant invertebrates:	-	AEMP Variable

Notes
[a] EEM Technical Guidance Document (Environment Canada 2012) and the MDMER (Government of Canada, 2002).
- = not applicable; AEMP = Aquatic Effects Monitoring Program; EEM = Environmental Effects Monitoring; MDMER = Metal and Diamond Mining Effluent Regulations.

Statistical Analysis

Statistical analyses will be conducted as per the EEM Technical Guidance Document (Environment Canada, 2012) and other approaches where warranted. Univariate (e.g., analysis of variance [ANOVA]) and multivariate statistical analysis techniques (e.g., nonmetric multidimensional scaling [nMDS], Mantels Test) may be used. If significant differences are observed between the exposure and reference areas, relationships between habitat variables and the benthic invertebrate metrics will be evaluated using tools such as calculating Spearman rank correlation coefficients and examining scatter plots. Statistical tests will be considered significant at a P-value ≤0.10, as recommended for EEM programs.

Univariate Analysis

With the exception of the Bray-Curtis Index, univariate statistical analyses will be undertaken to evaluate whether there are statistically significant differences in the benthic endpoints among sampling areas (i.e., Near-field, Mid-field, and Reference areas). Prior to statistical analysis, data will be evaluated for normal distribution and equality of variances to inform whether the data should be transformed and whether appropriate parametric (e.g., one way ANOVA) or non-parametric (e.g., Kruskal-Wallis one-way

ANOVA) tests should be employed. Selection of the appropriate parametric or non-parametric test will depend on applicability after reviewing the data and whether test assumptions are met. It should be noted that ANOVA is generally considered robust for detecting difference even if the data violate assumptions of normality.

The magnitude of differences between area means will be calculated for significantly different pairwise comparisons. The critical effect size (CES) will be calculated as plus or minus two standard deviations (±2 SD) of the reference area mean (Environment Canada, 2012). Magnitudes of differences between reference and the exposure areas will be considered biologically significant if they exceeded the CES.

Post hoc power analysis will be conducted for non-significant results to determine the actual power to detect an ecologically meaningful effect in the relevant endpoints.

Non-metric Multidimensional Scaling (nMDS)

To further assess differences in benthic community composition between sampling areas, community structure will also be summarized using the non-parametric ordination method of multidimensional scaling (Clarke, 1993). This ordination method allows visual identification of community-level differences among areas by representing abundance data in two or three dimensions. A Bray-Curtis resemblance matrix will be generated on log(x+1) data, and the nMDS procedure will be applied to this matrix where, using rank order information, the relative position of stations in terms of taxa abundances can be determined on an ordination plot. Goodness-of-fit will be determined by examining stress values. Lower stress values (i.e., less than 0.10) indicate a greater goodness-of-fit of ordination results to the input data, whereas higher stress values (i.e., greater than 0.20) must be interpreted with caution, and higher dimensions (i.e., 3-D) might be needed to describe the dataset (Clarke, 1993).

Assessment of Relationships with Habitat Variables

If warranted based on the magnitude of habitat variation, relationships between habitat variables and the benthic invertebrate endpoints will be evaluated using Spearman rank correlation coefficients and examining scatter plots. Habitat variables to be considered will include water depth, sediment grain size (e.g., percent fine sediments), and total organic carbon content, and potentially other variables. In addition, where appropriate, the findings of the benthic invertebrate data analysis will be further interpreted in light of results of other monitoring components, such as changes in sediment and water quality.

Comparison to FEIS Predictions

If the above analysis identifies a biologically significant difference between reference and exposure area benthic communities that is outside of the normal range, results will be evaluated further to determine whether the observed change in the benthic community is within FEIS predictions.

4.4.6 Quality Assurance and Quality Control

The QA/QC procedures employed in the collection, processing, and analysis of benthic invertebrate samples and supporting information will be consistent with the EEM Technical Guidance Document (Environment Canada, 2012).

Samples will be collected following standard sampling protocols by qualified personnel using appropriate sampling equipment. Quality control procedures will include estimating sample sorting efficiency and subsampling accuracy and precision, should subsampling be required. Ten percent of the samples will be re-sorted. A reference collection will be prepared, consisting of several representative specimens from each taxon. The reference collection will be archived with the taxonomist, for possible comparative purposes with benthic invertebrate community data from future studies and QC of future taxonomic identification.

Office-related QA will include using appropriately trained personnel for each task, senior review of work, standardized data handling/summary tools, and filing of original data. A second person will make quality checks of supporting data entered from field data sheets, spot checks of calculations performed during the data summary and analysis stage, and review of tables containing both summary data and statistical results.

4.5 Sediment Quality

4.5.1 Revisions in Version 3

No changes are proposed to the sediment chemistry monitoring program in Version 3 of the AEMP Design Plan.

4.5.2 Objectives

The objectives of the sediment quality monitoring program are:

- Verify predictions made in the FEIS in relation to sediment quality in Meliadine Lake,
- Characterize sediment quality,
- Collect supporting data for the benthic invertebrate and water quality components to aid interpretation of results (as per the MDMER), and
- Provide data to inform adaptive management intended to reduce or eliminate Mine-related effects to sediment quality in Meliadine Lake.

4.5.3 Study Design and Schedule

The sediment quality monitoring program is conducted every three years in mid-August. The sediment sampling stations are co-located with the benthic invertebrate stations at MEL-01, MEL-02, MEL-03, and MEL-05. Coordinates and water depth at each station are provided in **Table 4-1**.

4.5.4 Field and Laboratory Methods

Bottom sediment samples are collected to support the benthic invertebrate study according to methods outlined in the EEM Technical Guidance Document (Environment Canada, 2012) and sample collection methods specified by the accredited laboratories.

Sediment samples are collected using a petite Ponar (6” x 6”). The top 5 cm of the grab is retained for analysis, and material from up to five grabs will be combined and homogenized into a composite sample in the field. Physical descriptions of the sediment samples will be recorded, and photographs of representative samples taken. Samples will be collected in containers provided by an accredited analytical laboratory and shipped in coolers with ice-packs. The standard suite of parameters and target detection limits are provided in **Table 4-11**.

4.5.5 Data Analysis and Interpretation

Sediment data from the exposure areas will be evaluated by a multi-step process that focuses on comparing current sediment chemistry results in the exposure areas with data collected from the baseline period.

Comparisons to Sediment Quality Guidelines

Sediment quality data will be compared to applicable Canadian Sediment Quality Guidelines developed by the CCME will (i.e., ISQGs and probable effect levels [PELs]; CCME 1999, 2002). The ISQG is the concentration of a substance below which an adverse effect on aquatic life is unlikely, and the PEL is the concentration of a substance above which adverse effects are expected to occur frequently, but not always. In practice, the application of generic numeric guidelines has yielded a high percentage of false positives (Chapman and Mann, 1999). The observation of a sediment concentration above the PEL value for a given parameter should not be interpreted as an indication that actual ecological harm has occurred or will occur, but rather that this is a possibility.

Table 4-11. Sediment parameters and target detection limits

Physical Tests	Detection Limit	Metals (mg/kg dry weight)	Detection Limit
pH (1:2 soil:water)	0.1	Iron (Fe)	50
Particle Size (%)		Lead (Pb)	0.5
Cobbles (>3in.)	1	Lithium (Li)	2
Gravel (4.75mm - 3in.)	1	Magnesium (Mg)	20
Medium Sand (0.425mm - 2.0mm)	1	Manganese (Mn)	1
Fines (<0.075mm)	1	Mercury (Hg)	0.05
Coarse Sand (2.0mm - 4.75mm)	1	Molybdenum (Mo)	0.1
Fine Sand (0.075mm - 0.425mm)	1	Nickel (Ni)	0.5
Organic Carbon (%)		Phosphorus (P)	50
Total Organic Carbon	0.05	Potassium (K)	100
Metals (mg/kg dry weight)		Selenium (Se)	0.2
Aluminum (Al)	50	Silver (Ag)	0.1
Antimony (Sb)	0.1	Sodium (Na)	50
Arsenic (As)	0.1	Strontium (Sr)	0.5
Barium (Ba)	0.5	Sulfur (S)	1000
Beryllium (Be)	0.1	Thallium (Tl)	0.05
Bismuth (Bi)	0.2	Tin (Sn)	2
Boron (B)	5	Titanium (Ti)	1
Cadmium (Cd)	0.02	Tungsten (W)	0.5
Calcium (Ca)	50	Uranium (U)	0.05
Chromium (Cr)	0.5	Vanadium (V)	0.2
Cobalt (Co)	0.1	Zinc (Zn)	2
Copper (Cu)	0.5	Zirconium (Zr)	1

Temporal Trends

Version 1 of the AEMP Design Plan specified that the Normal Ranges for sediment chemistry would be used to provide context when interpreting changes in sediment chemistry (Golder, 2016). Normal range estimates presented in the 2018 AEMP pooled all reference and baseline data collected in Meliadine Lake instead of calculating a Normal Range for each basin (Golder, 2019). The concentrations of some metals often show considerable spatial heterogeneity in lakes located close to mineralized areas. Metals concentration in sediment are often naturally variable in lakes close to mineralized areas.

The relevant point of comparison is whether concentrations are changing within the near-field and mid-field areas over time, as opposed to assessing differences between the near-field, mid-field, and

reference areas. Changes in sediment chemistry over time within the exposure areas (MEL-01 and MEL-02) will be assessed statistically, for example with a before-after model that compares sediment chemistry data from the baseline period (pre-2018) to the current year. Before-after statistical models assumes that annual variability in sediment chemistry is negligible in absence of mining-related inputs.

4.5.6 Quality Assurance and Quality Control

Field Collection

Sample collection procedures described for the water sampling program are implemented to ensure the sediment chemistry data are reliable and accurate.

Laboratory QA/QC

Laboratory QA/QC procedures for sediment are described above for water in **Section 4.2.6**.

Field QA/QC

Field QA consists of taking care between sampling areas by rinsing and cleaning the sampling gear for sediment grabs (Petite Ponar grab, stainless steel compositing bowls and spoons) using site water and phosphate-free cleaning detergent, to avoid the possibility of cross-contamination.

Field QC measures include collection and analysis of at one field duplicate for every 10 samples (approximately). The DQOs for field duplicates are 1.5-times the laboratory RPDs. If the concentrations are less than 3-times the DL, the DQO is <1.5-times the difference between field duplicates.

4.6 Fish Health Study

4.6.1 Revisions in Version 3

The fish health study in Version 3 of the AEMP Design Plan matches the Cycle 3 EEM study design that was completed in August 2024. The program included lethal studies for Threespine Stickleback and Lake Trout. The Cycle 3 study design for both species was essentially a repeat of the study design used for the Cycle 2 EEM program in August 2021. The Technical Advisory Panel did not recommend any changes to the Threespine Stickleback or Lake Trout studies for the Cycle 3 EEM and the study designs are therefore considered acceptable for the AEMP.

4.6.2 Objectives

The objectives of the fish health component are as follows:

- Determine whether Mine effluent has an effect on fish populations in Meliadine Lake,
- Verify predictions made in the FEIS pertaining to fish health,

- Meet the requirements of the MDMER,
- Recommend appropriate changes to the fish health program for future years, and
- Provide data to inform adaptive management intended to reduce or eliminate Mine-related effects to fish health in Meliadine Lake.

4.6.3 Key Considerations for the Fish Health Study

Sentinel Species

The AEMP fish health study includes a small-bodied species (Threespine Stickleback) and a large-bodied species (Lake Trout). Within the scope of the EEM program, neither species is ideal for monitoring potential effects from exposure to effluent[10]. The main drawback for both species is effects to reproductive endpoints are difficult or impractical to assess. Threespine Stickleback are batch spawners, and at any given time the gonads of mature males and females can be either ripe or resting. The reproductive strategy for this species means gonad endpoints cannot be reliably assessed. For Lake Trout, relatively few mature Lake Trout spawn each year. For example, none of the males captured from Atulik Lake and Peter Lake in 2021 were ripe, and the numbers of ripe females in Meliadine Lake, Atulik Lake, and Peter Lake were 4, 1, and 3, respectively (Azimuth and Portt, 2022). To meet size requirements to detect effects for gonad endpoints, an unacceptably large number of fish would need to be lethally sampled.

Another disadvantage with using Lake Trout as large-bodied fish species for EEM is they have a large home range. Because they roam throughout Meliadine Lake, exposure to effluent is not continuous. The Technical Guidance Document recommends choosing sentinel species with small home ranges to increase the likelihood of exposure to effluent.

Despite the disadvantages with using Threespine Stickleback and Lake Trout as sentinel species, there are no viable alternatives based on the catch data from the 2021 field program (minnow traps in **Table 4-12**; gillnets in **Table 4-13**).

[10] Selection of sentinel species for EEM programs is discussed in detail in Section 3.3 of the EEM Technical Guidance Document. Some of the attributes of an ideal sentinel species include: (1) benthic-dwelling, (2) limited mobility relative to the size of the study area, (3) abundant and easy to catch.

Table 4-12. Species-specific catch summary from minnow traps from the Cycle 2 EEM in Meliadine Lake in August 2021.

Area	Total soak time (hrs)	Species-specific catch summary[a]						THST CPUE[b]
		THST	NNST	SLSC	LKTR	BURB	RNWH	
MEL-01	6268.2	533	9	1	0	0	0	0.085
MEL-03	13596.9	2143	327	14	6	6	0	0.158
MEL-04	13492.0	1512	204	8	6	18	2	0.112

[a] Species include: THST = Threespine Stickleback, NNST = Ninespine Stickleback, SLSC = Slimy Sculpin, LKTR = Lake Trout, BURB = Burbot, RNWH = Round Whitefish.
[b] CPUE = catch per unit effort = (fish/hr soak time)

Table 4-13. Species-specific catch summary from gill nets during Cycle 2 EEM in Meliadine, Peter, and Atulik lakes August 2021.

Lake	Total soak time (hrs)	Species-specific catch summary					Lake Trout CPUE[a]
		Lake Trout	Arctic Char	Whitefish	Cisco	Arctic Grayling	
Meliadine	38.2	67	0	2	7	0	1.75
Peter	78.6	45	5	0	16	1	0.48
Atulik	71.6	38	0	0	19	0	0.63

[a] CPUE = catch per unit effort = (fish/hr soak time)

Recent Fish Health Studies

Fish health studies were conducted in 2018, 2021, and most recently in 2024. The fish health assessment in 2018 included a lethal and non-lethal sampling program for Threespine Stickleback. The program was designed specifically to meet EEM sampling requirements and was not carried out to meet specific requirements of the AEMP (e.g., only one reference area was sampled [MEL-03] and the Lake Trout program was not completed). None of the endpoints exceeded their respective critical effect sizes. No changes to the EEM study design were recommend for subsequent EEM studies.

The second fish study in 2021 was completed to satisfy Water Licence (AEMP) and MDMER (EEM) monitoring requirements. Effort was made to harmonize the two programs where possible, but there were some notable differences between the AEMP and EEM. Those differences are summarized below:

Threespine Stickleback

- **AEMP**: The AEMP study was a lethal study that targeted unparasitized male and female fish in the exposure area (MEL-01) and two reference areas (MEL-03 and MEL-04). Unparasitized fish were sampled to control for the potential confounding effect of parasitism on survival (age), growth (size-at-age), and condition (body weight at length, relative liver size). Reproductive endpoints were not used to determine effects to Threespine Stickleback.
- **EEM**: The study design for Cycle 2 that was first submitted to ECCC was identical to the AEMP study described above except only one reference area was originally proposed (MEL-03; control-impact study design). However, ECCC recommended that the lethal study should also include parasitized Threespine Stickleback to determine if the conclusions about the health of the population depend on which portion of the population is targeted for monitoring. The EEM lethal study was completed as ECCC requested for parasitized and unparasitized males and females from all three study areas (MEL-01, MEL-03, MEL-04).
- **Key findings from the Cycle 2 EEM**: Threespine Stickleback endpoints were generally similar between exposure and reference areas regardless of parasite status. When differences were observed for a given endpoint, the direction of change was inconsistent between the exposure and reference areas (MEL-01 vs MEL-03 or MEL-04) and differences were occasionally observed between the two reference areas, MEL-03 and MEL-04 (Azimuth and Portt 2022). In conclusion, the Cycle 2 EEM results indicated that the status of parasitism was not an important factor when assessing the potential effects of effluent exposure on Threespine Stickleback in Meliadine Lake.

Lake Trout

- **AEMP**: The Lake Trout study in Version 1 of the AEMP Design Plan is a before-after study design where results from the operational phase (after 2018) are compared to results from the baseline period (2015). Before-after studies are effective at determining if a change has occurred, but they cannot discern if the change is related to mining activities or natural.
- **EEM**: Two external reference lakes were included in the Lake Trout study for the Cycle 2 EEM program. The reference areas were Peter Lake and Atulik Lake. The AEMP and EEM program shared the same Lake Trout data from the exposure area (MEL-01). Reproductive endpoints were not included in the analyses, so sexes were pooled to assess effects to survival, growth, and condition endpoints.
- **Key findings from the Cycle 2 EEM**: Lake Trout from Meliadine Lake were older and heavier on average compared to Lake Trout from the reference lakes. Weight-at-age was the only endpoint that exceeded the CES between Meliadine Lake and the reference lakes.

4.6.4 Study Design and Schedule

Threespine Stickleback

The Threespine Stickleback health study is a multiple-control impact study design with MEL-01 as the exposure area and MEL-03 and MEL-04 as the internal reference areas in Meliadine Lake. The study focuses on parasitized fish[11]. The program is conducted every three years in early to mid-August.

Unbaited gee-style minnow traps (1/4" square mesh; 9" x 16") are the most effective method for capturing Threespine Stickleback from shoreline areas in Meliadine Lake. Set date and time, lift date and time, water depth, substrate (dominant and sub-dominant), and the number of individuals captured of each species are recorded for each trap set. Non-target species are released and Threespine Stickleback retained for lethal sampling are transported to the field lab back at the Mine.

Specific conductance (µS/cm), pH, dissolved oxygen (mg/L and % saturation), and water temperature (°C) data are collected in the field within the exposure and reference areas.

The following data will be collected as part of the lethal Threespine Stickleback study:

- total length to the nearest mm;
- total weight (1% precision; e.g., to the nearest 0.01 g for fish that weigh > 1 g; 0.001 g for fish that weigh < 1 g);
- liver weight (to the nearest 0.0001 g);
- maturity status; and
- presence of external deformities, lesions, tumors, or parasites.

For mature Threespine Stickleback, sex, gonad condition (resting or ripe), and gonad weight (in grams to the nearest 0.001 g) will be recorded. Fecundity will be determined for ripe females either by counting all the eggs (if less than 100) or by dividing the total ovary weight by weight of individual eggs (minimum of 100 eggs).

Otoliths will also be collected from each captured Lake Trout to determine age. Extracted otoliths will be placed in envelopes labeled with the sampling area, date, species, and specimen number. Age will be estimated based on the number and annuli counted in whole otoliths using transmitted light and a stereo microscope. As a QA/QC measure, annuli will be counted by a second person for at least 10% of the otoliths. Age data will be used to examine the associated endpoints (e.g., size or weight at age).

Based on power analyses completed for the Cycle 3 EEM study design, 23 parasitized males were required to achieve the target power of 90% based on liver weight versus body weight. The minimum

[11] The rationale behind targeting parasitized fish was provided in the Cycle 3 EEM study design (Azimuth and Portt 2024).

number of females was 20 based on the same endpoint (Azimuth and Portt, 2024). The target sample size for the Cycle 3 EEM was 25 parasitized male and female fish from each area.

Lake Trout

The Lake Trout health study is a multiple-control impact study design with MEL-01 as the exposure area and Atulik Lake and Peter Lake as the external reference areas. The study is conducted every 3 years in early to mid-August.

Gill nets will be set in the exposure area within the extent of the 1% plume. If Lake Trout cannot be captured within this area in sufficient numbers, fish will be collected as close to the 1% effluent plume as practicable. Nets used in 2021 consisted of a gang of four North American standard large mesh gill nets (1.83 m x 24.7 m). Each standard net consisted of 8 panels of different mesh sizes (76 mm, 114 mm, 51 mm, 89 mm, 38 mm, 127 mm, 64 mm, and 102 mm). Specific conductance (µS/cm), pH, dissolved oxygen (mg/L and % saturation), and temperature (°C) will be determined in the vicinity of the gill net locations to confirm effluent presence and absence of stratification.

The geographic coordinates of each end of each net will be recorded, as will the depth and the date and time of deployment and retrieval. Set duration will be determined in the field based on local conditions, with the objective of meeting the sample size requirements while also minimizing the mortality of additional Lake Trout and incident catch. The number of individuals of each species captured in each net will be recorded.

The following information will be determined for each Lake Trout that is sampled lethally during the Cycle 3 EEM:

- presence of external deformities, lesions, tumors, or parasites;
- fork length in millimeters (to the nearest mm);
- total weight in grams (to within 1% of total weight);
- liver weight in grams (to the nearest 0.1 g);
- presence of internal deformities, lesions, tumors, or parasites; and
- maturity status (i.e., mature or immature).

For mature Lake Trout, sex, gonad condition (resting or ripe), and gonad weight (in grams to the nearest 0.1 g) will be recorded for each sex. Fecundity will be determined for ripe female Lake Trout according to the same approach described above for Threespine Stickleback. Gonad endpoints are not evaluated because only a small number of Lake Trout spawn each year, and achieving the target sample size would require sacrificing an unacceptably large number of fish.

Otoliths will also be collected from each Lake Trout from the lethal study for aging. The otoliths will be mounded whole on a glass slide, ground to the core on one side, flipped to adhere the core area to the

glass, and then ground to a thin section on the other side. Age will be estimated based on the number of annuli counted using transmitted light and a stereo microscope. As a QA/QC measure, annuli will be counted by a second person for at least 10% of the otoliths. Age data will be used to potentially examine the associated endpoints (e.g., size or weight at age), in case sample sizes provided sufficient power to adequately detect the effects.

Based on post-hoc power analyses completed for the Cycle 3 EEM study design, 26 Lake Trout (both sexes combined) are required to achieve target power of 0.9 based on the liver weight versus length endpoint (Azimuth and Portt, 2024).

4.6.5 Data Analysis for Lethal Surveys

Assessment and interpretation of the Lake Trout and Threespine Stickleback data will follow the approach outlined in the EEM Technical Guidance Document (Environment Canada, 2012) and recommendations from Technical Advisory Panel's review of the EEM study design. The workflow and analyses described below are for a standard lethal study.

Initial Data QA/QC

Data will be entered into spreadsheets and compared with original datasheets. Any errors or omissions that are identified will be corrected. Scatterplots of length versus weight will be prepared. If aberrant values are identified, original data sheets will be re-checked to ensure that these are not due to transcription errors. Any transcription errors found will be corrected. If clearly aberrant values for length or weight occur in the original data, these will be eliminated from the dataset.

Catch Data Summary

Catch-per-unit-effort provides an estimate of abundance by standardizing catch data according to fishing effort. For all fish captured during the health survey, catch-per-unit-effort will be calculated and summarized by area and sampling method to document the amount of effort expended to collect the required number of fish. Total numbers of fish collected and processed will be presented in summary tables by area.

Calculated Indices

Condition (K) will be calculated using the formula:

$$K = \frac{total\ weight}{total\ length^3} \times 100{,}000$$

Gonado-somatic index (GSI) will be calculated using the formula:

$$GSI = \frac{gonad\ weight}{total\ weight} \times 100$$

Liver somatic index (LSI) was calculated using the formula:

$$LSI = \frac{liver\ weight}{total\ weight} \times 100$$

Summary Statistics

Summary statistics (sample size, mean, median, minimum, maximum, standard deviation, standard error) will be calculated for each species and measurement endpoint evaluated in the study.

Length and Weight Distributions

Both length and weight distributions will be compared between sampling areas using pooled data. Skewness and kurtosis will be determined for both raw and $\log_{10}$ transformed distributions at each and divided by their respective standard errors. A value greater than two will be taken to indicate that a distribution deviates significantly from normal. As normality is an assumption of ANOVA, if either the raw or transformed data have values of skewness or kurtosis divided by their respective standard errors that are less than two, then the data will be analyzed using an ANOVA. Otherwise, the Kruskal-Wallace test will be used to compare distributions between areas.

Analysis of Covariance

ANCOVA is used to assess whether significant differences between the exposure and reference areas were present in the following relationships:

- total weight versus fork length,
- fork length versus age,
- total weight versus age,
- liver weight versus fork length, and
- liver weight versus total weight.

ANCOVA is used to test for significant differences in intercepts and slopes between the areas using $\log_{10}$ transformed values where appropriate. Significant differences are evaluated using alpha and beta equal to 0.1 (Environment Canada, 2012a). In cases where the interaction term was not significant (i.e., homogeneity of slopes between the exposure and reference areas), the reduced model was used to assess significance and effect sizes. In cases where the interaction term was significant, but accounted for <2% of the total variation in the response variable, the reduced model was considered appropriate

and used to assess significance and effect sizes as per Barrett et al. (2010). If differences in either slopes or intercepts existed, then pair-wise comparisons were used to determine which pairs differed.

Residuals from each ANCOVA were examined for normality and outliers. Observations producing large Studentized residuals (i.e., >4), if present, were removed from the dataset, and the analysis was repeated. Any changes in conclusions after removing outliers were carefully considered. This process was continued until no additional outliers were identified.

The percent difference in least-square means $(\bar{\chi})$ between the exposure area in Meliadine Lake and the reference lakes was calculated as:

$$\%Difference = \frac{\bar{\chi}_{exposure} - \bar{\chi}_{reference}}{\bar{\chi}_{reference}} \times 100$$

For log-transformed values, the least-square mean values are antilogs of the calculated values.

Table 4-14. Effect indicators, endpoints, and statistical tests used to determine effects to fish.

Effect Indicator	Endpoint	Dependent Variable	Covariate	Statistical Procedure	Critical Effect Size
Survival	Age	-	-	ANOVA	25%
Size	Length-frequency distribution	-	-	Kolmogorov-Smirnov Test	-
	Length	-	-	ANOVA	-
	Total Weight	-	-	ANOVA	-
Growth (Energy Use)	Size-at-age	Total Weight	-	ANOVA	25%
		Length	-	ANOVA	25%
Condition (Energy Storage)	Condition	Total Weight	Length	ANCOVA	10%
		Carcass Weight	Length	ANCOVA	10%
	Relative Liver Size	Liver Weight	Length	ANCOVA	25%
		Liver Weight	Carcass Weight	ANCOVA	25%

Statistical Analysis

Length, Weight, and Age Distributions

Length, weight, and age distributions will be compared between sampling areas for male and females. Skewness and kurtosis will be determined for both raw and $\log_{10}$ transformed distributions at each and divided by their respective standard errors. A value greater than two will be taken to indicate that a distribution deviates significantly from normal. As normality is an assumption of ANOVA, if either the raw or transformed data have values of skewness or kurtosis divided by their respective standard errors

that are less than two, then the data will be analyzed using an ANOVA. Otherwise, the Kruskal-Wallace test will be used to compare distributions between areas.

Weight and length versus age

Given that ages are likely to span four years or less and that some ages will be poorly represented, size at age will be compared for ages that are well-represented using ANOVA or, if warranted due to violation of assumptions, the Mann-Whitney test.

Analysis of Covariance

ANCOVA will be used to determine if significant differences between the exposure and reference area occur in the following relationships:

- total weight versus total length,
- liver weight versus total weight, and
- gonad weight versus total weight.

Using $\log_{10}$ transformed values where appropriate, ANCOVA will be used to test for significant differences in intercepts and slopes between the areas. Significant differences will be evaluated using an alpha (α) of 0.1 (Environment Canada, 2012a). In cases where the interaction term is not significant (i.e., homogeneity of slopes between the exposure and reference area), the reduced model will be used to assess significance and effect sizes. In cases where the interaction term is significant, but accounts for <2% of the total variation in the response variable, the reduced model will be considered appropriate and used to assess significance and effect sizes as per Barrett et al. (2010).

Residuals from each ANCOVA will be examined for normality and outliers. Observations producing large Studentized residuals (i.e., >4) will be removed from the dataset, and the analysis will be repeated. Any changes in conclusions will be considered. This process will be continued until no additional outliers are identified.

The percent difference in least-square means $(\bar{\chi})$ between the exposure and reference areas in Meliadine Lake will be calculated as:

$$\%Difference = \frac{\bar{\chi}_{exposure} - \bar{\chi}_{reference}}{\bar{\chi}_{reference}} \times 100$$

When log transformed data are analyzed, the least-square mean values used will be antilogs of the calculated values.

Power Analysis

Power analysis was used to determine, *a posteriori*, the probability of detecting a 10% (weight versus length) or 25% (gonad weight versus total weight, liver weight versus total weight) increase in the

parameters of interest, assuming a 10% probability of committing a Type I error, and given the sample sizes, mean values, and the unexplained variability (i.e. the population standard deviation) from this study. Power was calculated by re-arranging the following power equation (Green, 1989):

$$n = \frac{1.5(t_{\alpha} + t_{\beta})^2 \sigma^2}{\delta^2}$$

Where:

n is the number of fish,

σ is the population standard deviation,

δ is the specified effect size,

t_{α} is the Student's *t* statistic for a two-tailed test with significance level α,

t_{β} is the Student's *t* statistic for a one-tailed test with significance level β.

In cases where no significant differences are observed in effect endpoints, *post-hoc* power analyses will be performed to determine if there was sufficient power to detect differences equivalent to the respective CES in the population.

4.6.6 Quality Assurance/Quality Control

The QA/QC procedures are designed such that field sampling, laboratory analyses, data entry, data analyses, and report preparation produce technically sound and scientifically defensible results. As part of routine QA/QC for field operations, equipment (e.g., water quality meters, weigh scales) will be calibrated and samples will be collected by experienced personnel and will be labelled, preserved, and shipped according to standard protocols. Specific work instructions outlining each field task in detail will be provided to the field personnel by the task manager and reviewed prior to the start of the field program.

Field notes will be recorded in waterproof field books and on pre-printed waterproof field data sheets in either pencil or indelible ink. Data sheets and all sample labels will be checked at the end of each field day for completeness and accuracy. Chain-of-custody forms will be used to track the shipment of all samples. For aging structures, 10% of the prepared sections will be re-aged by an independent fish ageing specialist. If there is a discrepancy greater than 10% between the specialist's results and the initial results, all samples will be re-analyzed. For every ten fecundity samples, one sample will be recounted by a second person. If the re-count of the sample is within 10% of the initial count, the initial count will be regarded as acceptable and no re-count of the remaining samples will be required. If the re-count is not within 10% of the initial count, the initial count will be regarded as unacceptable and the remaining nine samples will be re-counted. The QA/QC procedure will be repeated until re-counts are within 10% of the previous count.

The QA/QC for data entry involves checking a minimum of 10% of the data for data entry errors, transcription errors, and invalid data. This checking will be done by an independent person from the person who entered the data. If an error is found, every datum will be checked. Statistical results will be independently reviewed by a qualified senior biologist. Tables containing summary data and statistical results will be reviewed and values verified by a second person.

4.7 Fish Tissue Chemistry

The Lake Trout tissue chemistry monitoring program was included in the AEMP primarily to verify that the Mine is not contributing to changes in tissue chemistry that would affect the useability of the fishery for traditional and recreational purposes. Threespine Stickleback were included in the study design to characterize bioaccumulation and trophic transfer of contaminants through the food web (i.e., link between the lower trophic levels and predatory fish species).

The combined effect of warmer temperatures and increased precipitation were cited in the 2014 FEIS as potential factors that could lead to higher concentrations of metals in Arctic fish species (Carrie et al., 2010; Barletta et al., 2012; Dijkstra et al., 2013). However, the overall conclusion was that the effect of mining activities on fish, including changes in tissue chemistry, would be negligible compared to the spatial and temporal scale of climate-related changes.

4.7.1 Revisions in Version 3

In Version 1 of the AEMP Design Plan, Lake Trout muscle, liver, and kidney samples were collected and submitted for metals analysis. Liver and kidney samples are included to help interpret the results of the Lake Trout health assessment if adverse effects to survival, energy use, and/or energy storage are identified. For efficient use of resources, the liver and/or kidney samples will be analyzed only if results from the Lake Trout health assessment indicate there are adverse mining-related effects to Lake Trout in Meliadine Lake.

4.7.2 Objectives

The objectives of the fish tissue chemistry component are as follows:

- Determine if effluent is causing an increase in metal concentrations in fish tissue in Meliadine Lake, including whether fish tissue chemistry has been altered in such a way as to limit fish use by humans,
- Verify predictions made in the FEIS pertaining to fish tissue metal concentrations,
- Meet the requirements of the MDMER,
- Aid in the interpretation of the fish health study,
- Recommend appropriate changes to the fish tissue chemistry program for future years, and

- Provide data to inform adaptive management intended to reduce or eliminate Mine-related effects to fish tissue chemistry in Meliadine Lake.

These objectives for fish tissue chemistry are addressed through the following key question:

- Are tissue metal concentrations in fish from Meliadine Lake exposure areas increasing due to mining activities?

4.7.3 Study Design and Schedule

Fish tissue chemistry will be collected from Threespine Stickleback (carcasses) and Lake Trout (muscle) every three years coinciding with the fish health study. Historical samples sizes for each species, area, and tissue type are summarized in **Table 4-15**. Sample sizes are subject to change as additional data is collected to understand the variability within and between areas.

Table 4-15. Overview of the fish tissue sampling programs for the AEMP.

Species	Year	Lake/Area	Area Status	Phase	Sample Sizes			
					Muscle	Liver	Kidney	Carcass
Lake Trout	1998	Meliadine Lake	Control	Baseline	34	34	33	-
	2015	Meliadine Lake	Control	Baseline	60	60	60	-
	2021	Meliadine Lake	Impact	Operations	42	42	42	-
		Atulik Lake	Control	Reference	24	0	0	-
		Peter Lake	Control	Reference	24	0	0	-
Threespine Stickleback	2015	MEL-01	Control	Baseline	-	-	-	60
	2017	MEL-03	Control	Reference	-	-	-	67
		MEL-04	Control	Reference	-	-	-	67
	2021	MEL-01	Impact	Operations	-	-	-	40
		MEL-03	Control	Reference	-	-	-	40
		MEL-04	Control	Reference	-	-	-	40

4.7.4 Field and Lab Methods

A subset of the Threespine Stickleback and Lake Trout from the populations study will be processed for tissue metals analysis. For the Threespine Stickleback study, specimens will be chosen based on similar size classes among the study areas and, ideally, from the size class from previous cycles. Fish in this size range are typically three to four years old. Prior to analysis, the lab will be consulted to verify that the mass of individual fish is sufficient to meet the target detection limits (**Table 4-16**).

Table 4-16. Parameters and target detection limits (mg/kg wet weight) for fish tissue analysis.

Target Detection Limits (mg/kg wet weight) [a]							
Aluminum	0.4	Cesium	0.001	Mercury	0.001	Strontium	0.01
Antimony	0.002	Chromium	0.01	Molybdenum	0.004	Tellurium	0.004
Arsenic	0.004	Cobalt	0.004	Nickel	0.04	Thallium	0.0004
Barium	0.01	Copper	0.02	Phosphorus	2	Tin	0.02
Beryllium	0.002	Iron	0.6	Potassium	4	Titanium	0.05
Bismuth	0.002	Lead	0.004	Rubidium	0.01	Uranium	0.0004
Boron	0.2	Lithium	0.1	Selenium	0.01	Vanadium	0.02
Cadmium	0.001	Magnesium	0.4	Silver	0.001	Zinc	0.1
Calcium	4	Manganese	0.01	Sodium	4	Zirconium	0.04

Notes:
The detection limits are from the 2021 AEMP.

For Lake Trout, subsamples of muscle, liver and kidney tissue will be collected from all of the lethally sampled fish in each area. The muscle samples will be submitted for analysis, with a target sample size in each area of 20 to 30 fish (pooled sexes). The liver and kidney samples will be archived. Metals analysis of kidney and/or liver samples may be undertaken on some or all of the samples to help interpret the results of the fish population study.

Field tools will be cleaned between dissections to minimize the potential for cross contamination between samples, or new disposable tools will be used for each fish (e.g., scalpels). Tissue samples will be weighed, packaged, and labelled with the appropriate fish identification number.

4.7.5 Data Analysis and Interpretation

Descriptive statistics (i.e., sample size, mean, standard deviation, standard error, minimum, and maximum) and statistical comparisons will be presented in an appendix for all metals concentrations.

Data analysis will focus on comparing concentrations among the exposure and reference areas and over time. Parameters that are detected in less than 50 % of the samples from the Meliadine Lake exposure areas in the current year will not be carried forward for statistical analysis.

Spatial and temporal patterns for the Threespine Stickleback data will be assessed using analysis of variance (ANOVA) and pair-wise comparisons (Tukey's honestly significant difference test) among the area-year combinations.

Lake Trout have longer lifespans than small-bodied fish species like Threespine Stickleback, and progressively accumulate bioaccumulative metals such as mercury and selenium in their tissue over time. This can lead to size-related differences in tissue concentrations, which can lead to biased results if

the underlying size-metal relationships are not considered. ANCOVA explicitly considers the influence of size-related covariates (e.g., length, weight, or age) when testing for differences in tissue metals concentrations between or among years. ANCOVA analysis will be conducted with length as the covariate according to approach outlined in the Lake Trout health assessment (**Section 4.6.5**).

4.7.6 Quality Assurance and Quality Control

The analytical laboratory will analyze a series of sample blanks, spikes, and laboratory duplicates, and certified reference standards (CRMs) will be run in parallel with the tissue chemistry samples. The results of these internal QA/QC processes will be reported with the laboratory data and any deviations from acceptable data quality objectives will be reported. If acceptable limits are exceeded, samples will be re-assessed and, if necessary and possible, re-analyzed.

Laboratory data will be screened in a manner similar to the water quality data (**Section 4.2.6**). Data entry will be reviewed to verify completeness (e.g., no data entry errors, transcription errors, and invalid data). Statistical test results will be independently reviewed by a second, competent statistician. Tables containing both summary data and statistical results will be reviewed and values verified by a second, independent individual.

5 PENINSULA LAKES STUDY

5.1 Overview

5.1.1 Environmental Setting

Several small watersheds drain to Meliadine Lake from the peninsula between the south and east basins of Meliadine Lake. The peninsula watersheds comprise an extensive network of small lakes, ponds, and interconnecting streams. The lakes within the peninsula are generally small (<90 ha in area) and shallow (between 2 and 5 m in maximum depth). They do not freeze to the bottom. They are connected to each other (and to Meliadine Lake) through short stream sections; however, they can often be isolated by limited flow during the summer/fall and frozen stream conditions during the winter.

Lakes on the peninsula were characterized as well-oxygenated, with pH values indicative of slightly basic conditions, low sensitivity to acid deposition, and low to moderate ionic strength during the baseline period. Parameter concentrations were typically below relevant guidelines. Sediment samples from the Peninsula Lakes were a mix of sand and fine sediments with concentrations of some metals above CCME interim sediment quality guidelines (ISQG) values (e.g., arsenic, chromium, and copper), which is similar to Meliadine Lake under baseline conditions.

5.1.2 Study Areas

Three lakes were selected for water quality monitoring as part of the AEMP: Lakes A8, B7, and D7. These are headwater lakes in three different peninsula watersheds. Lake B7 and Lake A8 are close to major mine infrastructure while Lake D7 is located to the west in a watershed that was not directly impacted by development of the Mine. Lakes A8 and D7 were used by Inuit for collecting small Lake Trout and Lake Whitefish (Wesley from Rankin Inlet Hunters and Trappers Organization). Surface area, depth, and shoreline information for each lake is provided in **Table 5-1**.

Table 5-1. Morphological Characteristics of AEMP Peninsula Lakes

Lake	Surface Area (ha)	Volume ($m^3 \times 10^3$)	Depth(m)		Total Shoreline Length (km)
			Mean	Maximum	
A8 (former)	89.7	1,419.3	1.6	4.2	7.5
B7	58.1	852.5	1.5	5.1	5.5[a]
D7	72.5	1,183.4	2.8	5.2	5.2
E3 (new)	56.8	880.9	1.6	4.7	Not reported

Source: Golder (2012a) Aquatic Baseline Synthesis Report.
[a] Includes shoreline length around two islands.

5.2 Water Quality

5.2.1 Revisions in Version 3

As mentioned in the introduction, Agnico Eagle has been granted approval to mine deposits that are located in the vicinity of Lake A8 (Wesmeg and Pump **Figure 2-3**). To develop these gold deposits, Lake A8 will need to be dewatered. Agnico Eagle is evaluating potential lakes to include in the Peninsula Lakes study to monitor changes in water quality caused by development of these deposits, as well as F-Zone.

Starting in 2025, Lake A8 will be removed from the study design. Agnico Eagle recommends adding Lake E3 to the study design. Of the lakes to add to the study design, Lake E3 is the most suitable option based on its size (one of the larger lakes near the Mine with overwintering habitat [**Table 5-1**]), proximity to major mine infrastructure, and because water sampling has been conducted annually during the open water season since 2018.

5.2.2 Objectives

The primary objectives of the water quality component for the Peninsula Lakes study are as follows:

- Characterize and interpret water quality in the selected monitoring lakes for purposes of identifying effects related to the Mine,
- Verify and update the FEIS predictions and other submissions to the NWB, as applicable, relating to water quality,
- Assess the efficacy of impact mitigation strategies to minimize impacts to water quality,
- Provide data to inform management decisions to reduce or eliminate mine-related effects to water quality in the Peninsula Lakes, and
- If necessary, recommend changes to the water quality component of the AEMP for future years.

These objectives are addressed through the following key questions:

- Is water quality consistent with predictions outlined in the FEIS and less than AEMP Action Levels?
- Has water quality changed over time, relative to baseline conditions?

5.2.3 Sampling Design and Schedule

Water samples are collected from each lake in July and August from three fixed monitoring locations in Lake B7 and Lake D7. The Water Licence states that biannual water sampling is required at Lake E3 (MEL-15) during the open water season (shoreline sample; **Figure 2-2**). For the AEMP, surface water sampling in Lake E3 will be conducted in July and August to align with the timing of water sampling in Lake D7 and Lake B7.

5.2.4 Field and Lab Methods

The water quality monitoring program for the Peninsula Lakes study is based on the same methods outlined for Meliadine Lake. Limnology measurements are taken at each station in Lake B7 and Lake D7 along with a surface water sample from mid-depth in the water column. Grab samples are collected from the shoreline of Lake E3 for the Water Licence. To maintain consistency across years, shoreline grabs are recommended for Lake E3 moving forward.

Water samples are analyzed for the same parameters as the Meliadine Lake study (**Table 4-6**).

5.2.5 Data Analysis and Interpretation

Analysis and interpretation of the Peninsula Lakes water quality data includes screening the data against the AEMP Benchmarks/Action Levels and the Normal Range of baseline conditions. Other quantitative statistical techniques may be adopted to discern changes in water quality caused by mining activities compared to natural variability.

5.2.6 Quality Assurance and Quality Control

QA/QC procedures will be consistent with those described for water quality monitoring in Meliadine Lake (**Section 4.2.6**).

5.3 Biological Monitoring in the Peninsula Lakes

Biological studies may be included in future monitoring cycles if results of the water quality program indicate there are changes that could potentially impact the health of aquatic life.

6 RESPONSE FRAMEWORK

The AEMP Response Framework links monitoring results to management actions to maintain the assessment endpoints within acceptable ranges. It is a systematic approach to adaptive management, ensuring that environmental monitoring results trigger appropriate actions to mitigate potential impacts to the aquatic environment. This is accomplished by continually evaluating monitoring data and implementing follow-up actions (e.g., confirmation, further study, mitigation) at pre-defined levels of change in measurement endpoints (i.e., Action Levels).

Action Levels (i.e., Low, Moderate, and High) will be used within the Response Framework to determine if follow-up action is required to manage and reverse any detected changes in the aquatic environment. If a Low Action Level is reached for one or more components of the AEMP, a response action will be initiated. Specific terms used in the Response Framework include: Benchmarks, Action Levels, and Significance Threshold, and are defined as follows:

- **AEMP Benchmark**. the aquatic life guidelines (e.g., CCME or Federal Environmental Quality Guidelines), generic drinking water guidelines, or site-specific water quality guidelines (SSWQOs) used to screen the water chemistry data. As an added level of protection, early warning 'triggers' (equal to 75 % of the AEMP Benchmark) are used to identify parameters that are trending higher. This ensures that corrective action is taken before the exceedance of an AEMP Benchmark.
- **Action Levels**. Low, Moderate, and High Action Levels are pre-defined levels of environmental change. They are often linked to benchmarks, results of statistical tests, or a combination of the two. A Low Action Level exceedance serves as an early-warning indication of the potential for adverse effects on an ecosystem component. Exceedance of a Low Action Level indicates a measurable change has occurred, but the magnitude is below the Significance Threshold. Moderate and High Action Levels are designed to identify measurable effects that are trending towards the Significance Threshold, and may trigger follow-up management actions or responses to slow, stop, and reverse the trend.
- **Significance Threshold**: a level of change that would result in significant adverse effects to key values of the environment that are to be protected. This is considered an unacceptable level of change or 'no go condition'. Significance Thresholds are based on the assessment endpoints. Failure to meet the assessment endpoints (e.g., suitability of water to support an aquatic ecosystem) would result in the Significance Threshold being met.

If a change in the monitoring data is detected that exceeds a Low Action Level, the best course of action will depend upon the type of effect observed. Examples of response actions are provided in **Table 6-1**.

Figure 6-1. Overview of the Aquatic Effects Monitoring Program Response Framework.

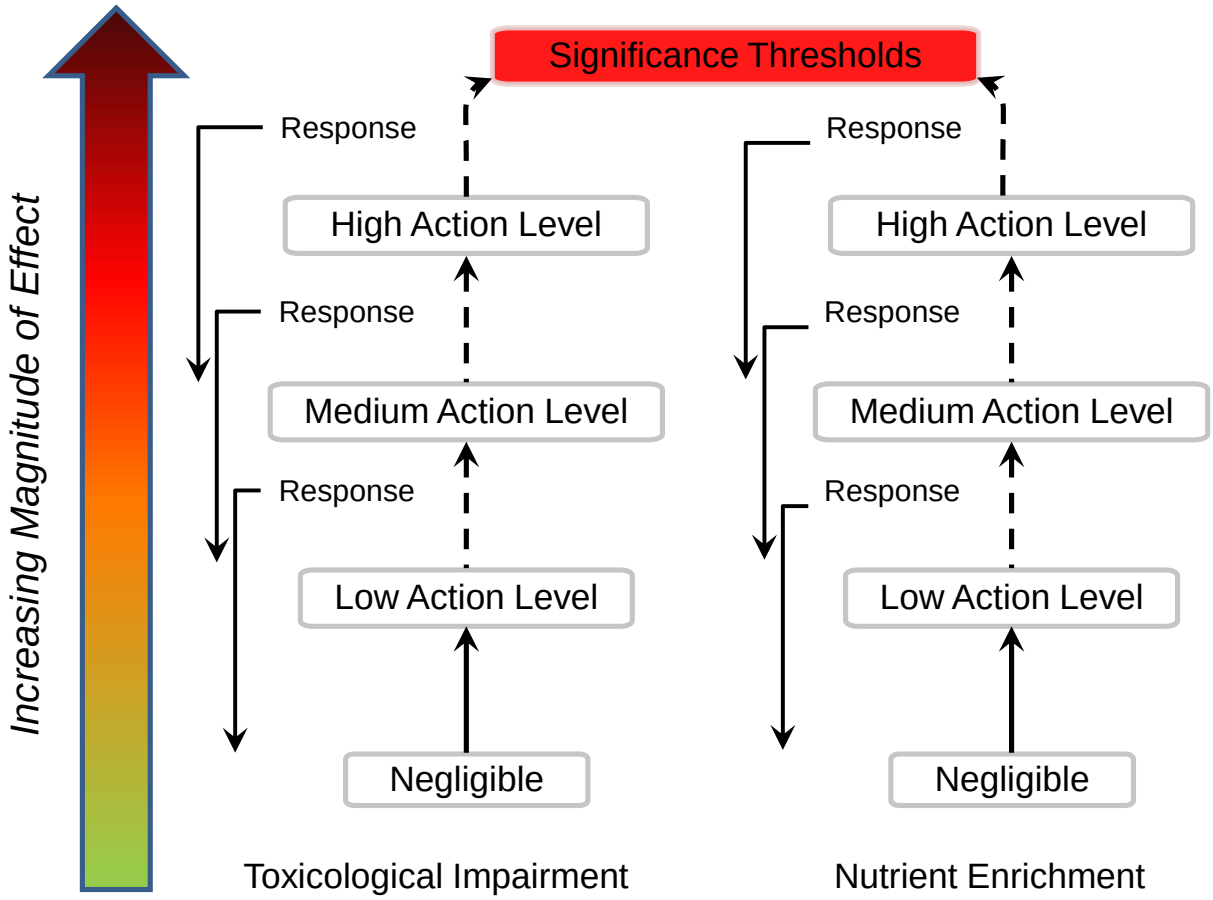

Table 6-1. Examples of Action Levels and Responses for Water Chemistry

Action Level	Example of Action Level to Support Impact Hypothesis "Toxicological Impairment"	Example of Action Level Response
Negligible[a]	no difference between reference and exposure areas or from baseline conditions; values of measurements endpoints within Normal Ranges	(none required)
Low	difference between reference and exposure areas, but below an applicable benchmark increasing trend toward conditions outside of Normal Range, or toward a benchmark	AEMP best practices Increase monitoring (e.g., establish new stations if the plume appears to be moving faster and farther than expected (e.g., establish new stations in the "narrows" between the Near-field and Mid-field) Confirm Low Action Level trigger Compare to FEIS predictions Investigate further to identify contributing factors from the Mine Examine ecological relevance Identify potential mitigation options Re-evaluate benchmark and revise if necessary Set Moderate and High Action Levels
Moderate	significant difference between reference and exposure areas, and benchmark exceeded consistently increasing trend approaching benchmark exceedance	AEMP best practices Notify NWB Confirm Moderate Action Level trigger Compare to FEIS predictions Prepare a response plan Investigate further to identify contributing factors from the Mine Examine ecological relevance and implications Implement mitigation and examine effectiveness of mitigation Update monitoring design
High	benchmarks consistently exceeded, or effect is above predictions but below the Significance Threshold[b]	AEMP best practices Notify NWB Confirm High Action Level trigger Compare to FEIS predictions Prepare a response plan Identify and implement improved mitigation to reverse trend Remediate

Notes:

AEMP Best Practices: evaluate causation/linkage to the proposed Mine, examine trends, predict trends where appropriate, examine linkage between exposure, toxicity, and field biological responses, examine ecological significance, confirm that benchmarks are appropriate and revise if warranted.

[a] Not an Action Level but is listed to provide an indication of the estimated magnitude of background variation.

[b] Significance Threshold is defined as the point at which an environmental change would be considered significantly adverse. The adaptive management actions are used to prevent a Significance Threshold from being reached.

6.1 Significance Thresholds

Significance Thresholds focus on key values to protect rather than the numeric values set as Action Levels. The Significance Thresholds span all monitoring components and both impact hypotheses (toxicological impairment and nutrient enrichment). They are the "no-go" condition for the Mine. The proposed Significance Thresholds include the following key "values" that are to be protected:

- water is safe for human and wildlife consumption,
- fish are safe for human and wildlife consumption, and
- the ecological function of the aquatic environment is maintained (i.e., there is adequate food for fish, and fish are able to survive, grow, and reproduce).

Based on these values, Significance Thresholds proposed for the AEMP are as follows:

- Water is not drinkable (human health and/or wildlife risk):
  - Safety of water for consumption will be considered through a human health and/or wildlife risk assessment for drinking water.
- Fish are not safe for consumption (human health and/or wildlife risk):
  - No contaminants of potential concern (COPCs) were identified in fish tissue in Meliadine Lake; this pathway was considered to be incomplete and was not retained for further assessment in the HHRA (Volume 10, 2014 FEIS). Risk assessment tools may be considered if the concentrations of COPCs in fish tissue are statistically significantly higher than other lakes in the region. Mercury is a special case because concentrations often exceed the Health Canada mercury consumption limit of 0.5 mg/kg wet weight because of the propensity for mercury to bioaccumulate in large, old lake trout in northern aquatic ecosystems.
- Ecological Function is not maintained:
  - Inadequate food for fish, fish are unable to survive, grow, or reproduce, and/or sustained absence of a fish species.

6.2 Low Action Level Assessment for Meliadine Lake

The Low Action Level Assessment for Meliadine Lake provides advanced warning of potential adverse effects to fish and other aquatic receptors from toxicological impairment (**Table 6-2**) and nutrient enrichment (**Table 6-3**). The assessment criteria were designed so that if a Low Action Level is exceeded, the results are reported, documented, investigated, and ultimately addressed (i.e., mitigation or operational changes are implemented) before Significance Thresholds would ever be reached. If a Low Action Level is reached, Medium and High Action Levels (with response actions) will be developed to support adaptive management.

Table 6-2. Low Action Levels for Toxicological Impairment for Meliadine Lake

Component	Assessment	Low Action Level Assessment Criteria[a]
Water Quality	End of Pipe Toxicity	Confirmed sublethal toxic effects on test organisms other than fish in end-of-pipe samples AND No sublethal toxic effects on fish in end-of-pipe samples
	Aquatic Life	Near-field mean above the Normal Range AND Statistically significant higher concentration in the Near-field compared to Reference AND Near-field mean exceeds 75 % of an AEMP Benchmark
	Human Consumption	Statistically significant higher concentration in the Near-field area compared to Reference AND Drinking water parameters in exposure area above 75 % of Health Canada's human health drinking water quality guideline (maximum acceptable concentration)
Phytoplankton	Aquatic Life	Phytoplankton community metrics at the Near-field area outside the range of baseline/reference conditions AND Change in direction and magnitude that are indicative of toxicological impairment
Benthic Invertebrates	Aquatic Life	Statistically significant difference in Near-field total density or richness compared to Reference AND Change in direction and magnitude indicative of toxicological impairment AND Difference in invertebrate density or richness with magnitude ≥CES[b] between reference and exposure areas
Fish Health	Aquatic Life	Statistically significant differences in fish health endpoints[c] between Near-field and Reference AND Change in direction and magnitude indicative of impairment of fish health AND Magnitude of effect above the CES[c]
Fish Usability	Human Consumption	Statistically significant difference in metal concentrations relative to reference AND Mean metal concentrations above a fish consumption guideline that is protective of human health

Notes:
[a] Only Low Action Levels are developed initially; Moderate and High Action Levels will be developed if the Low Action Level is reached.
[b] Critical effect size (CES) for benthic invertebrate community is two standard deviations of the current monitoring year's reference area data.
[c] Refer to **Table 4-14** for the fish health endpoints and corresponding critical effect sizes.

Table 6-3. Low Action Levels for Nutrient Enrichment for Meliadine Lake

Component	Assessment	Low Action Level Assessment Criteria[a]
Water Quality	Aquatic Life	Concentrations of total phosphorus (TP) in the Near-field area above the Normal Range, supported by temporal trends AND A statistically significant relative difference between the Near-field area and Reference for TP AND Average TP concentration in the Near-field area that exceeds 75 % of AEMP Benchmark
Phytoplankton	Aquatic Life	Near-field mean for total phytoplankton biomass above the upper bound of the Normal Range AND Change in direction and magnitude indicative of nutrient enrichment
Benthic Invertebrates	Aquatic Life	Statistically significant difference in total density or richness between Near-field and Reference Areas AND Change in direction and magnitude indicative of nutrient enrichment AND Difference in invertebrate density or richness with magnitude ≥CES[b] between reference and exposure areas
Fish	Aquatic Life	Statistically significant differences in fish health endpoints[c] AND Changes is in direction and magnitude that are indicative of nutrient enrichment AND Magnitude of effect above the CES[c]

Notes:
[a] Only Low Action Levels are developed initially; Moderate and High Action Levels will be developed if the Low Action Level is reached.
[b] Critical effect size for benthic invertebrate community will be two standard deviations of the current monitoring year's reference area data.
[c] Refer to **Table 4-14** for the fish health endpoints and corresponding critical effect sizes.

6.3 Peninsula Lakes Water Quality and Adaptive Management

Water quality data from the Peninsula Lakes are evaluated using the same approach as the Meliadine Lake study, including comparisons to (1) baseline conditions (Normal Range assessment), (2) water quality guidelines, and (3) predictions in the 2014 FEIS (if available). The objective is to ensure changes in water quality are detected early to mitigate against adverse effects to aquatic life. Water quality data from the Peninsula Lakes monitoring program are integrated with water quality data from compliance monitoring under the Water Licence (e.g., MEL-15, -16, -17, and -18) as part of a holistic approach to adaptive management within the Annual Report.

6.4 Plan Effectiveness

The AEMP is a clear and defensible monitoring design that complies with relevant laws and regulations. Through annual reporting, it verifies the efficacy of mitigation and management measures to prevent adverse effects on the freshwater receiving environment. Agnico Eagle may periodically evaluate the efficacy of monitoring, mitigation, and management activities using methods such as power analysis or time series analysis. This plan will be updated as needed if new and relevant monitoring methods become available.

7 REPORTING

Per Part B Item 2 of the Water Licence, an Annual Report must be submitted to NWB no later than March 31st of every year. Per Schedule B Item 17 of the Water Licence, this Annual Report must include the results of monitoring related to the AEMP. These results will be presented in an AEMP Report, which will be an attachment to the main Annual Report. The AEMP Report will include:

- A summary of Project activities during the monitoring interval.
- A summary of the monitoring data obtained during the most recent reporting period.
- Description of the methods used for data collection and analysis.
- Evaluation of Project-related effects on the measurement endpoints.
- Results of the Action Level assessment.
- Recommendations (e.g., additional sampling or analysis, adaptive management).

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APPENDICES

APPENDIX A
RECOMMENDATIONS, CONDITIONS, AND COMMITMENTS RELATED TO THE AQUATIC EFFECTS MONITORING PROGRAM

The Mine underwent an environmental assessment with the Nunavut Impact Review Board (NIRB) and a Type A Water Licence application process. A series of recommendations and conditions were listed In the NIRB decision report (NIRB, 2014). In addition, Agnico Eagle Mines Limited (Agnico Eagle) committed to a series of recommendations raised by various interveners during both the environmental assessment and the Water Licence process. A summary of the recommendations and conditions, and commitments made by Agnico Eagle to interveners during the regulatory process, which are directly relevant to the AEMP, are provided in **Table A-1**.

Table A-1. Recommendations, Conditions, and Commitments Related to the Aquatic Effects Monitoring Program

Commitment Number and Source	Recommendation / Condition / Commitment Details	Reference
Environmental Assessment		
NIRB Decision Report (NIRB 2014) Condition 30	The Proponent shall update its AEMP to include, at a minimum: Details for additional reference lakes to be included within its sampling and monitoring programs; Updates to include sedimentation within relevant monitoring programs; and Results from additional testing for mercury in fish tissue, and include test results in updated baseline data.	Reference Area: **Section 4.1.2** Sedimentation: not included in the AEMP Design Plan Mercury: Golder (2018)
FEIS KIA-IR-06	Agnico Eagle will engage the Inuit to ensure their assessment of whether the "Opportunity for traditional and non-traditional use" has been impaired.	**Section 1.4**
FEIS KIA-IR-11	Agnico Eagle will monitor water quality in the receiving environment to enable the identification of trends and additional adaptive management strategies, if required, including potential sediment and erosion control.	Meliadine Lake: **Section 4.2.5**
FEIS KIA-IR-22	The KIA are concerned about dissolved oxygen concentrations during vulnerable times of the year (i.e., low flow or under-ice). They recommended modelling of under-ice dissolved oxygen in the mixing zone. Agnico Eagle commits to monitoring under-ice dissolved oxygen concentrations in the mixing zone of Meliadine Lake.	DO modelling: FEIS Appendix 7.4A (Agnico Eagle, 2015) DO under ice: **Section 4.2.4**
FEIS KIA-IR-29	Agnico Eagle will conduct a survey to collect fish tissue chemistry to provide a recent baseline dataset.	Baseline fish tissue chemistry in Golder (2018) & **Section 4.7**
FEIS KIA-IR-NEW-08	KIA are concerned that water quality downstream in Peter Lake (downstream of the northwest outlet of Meliadine Lake) could be impacted, and have recommended a monitoring location in the Diana River watershed.	Agnico Eagle committed to monitoring water quality in Meliadine Lake near the northwest outlet (MEL-04) as an early warning to potential far downstream effects.
FEIS KIA-IR-NEW-09	For the purposes of future water quality monitoring programs, the term "differing from baseline" will be defined through calculations of normal range.	**Section 4.2.5**
FEIS KIA-IR-NEW-11	Agnico Eagle will assess the impact of Mine activities in part through the changes observed in the benthic macroinvertebrate community composition and density.	**Section 4.4**

Table A-1. Recommendations, Conditions, and Commitments Related to the Aquatic Effects Monitoring Program

Commitment Number and Source	Recommendation / Condition / Commitment Details	Reference
FEIS KIA-IR-NEW-12	Agnico Eagle has committed to analyzing tissue from fish in Meliadine Lake and select peninsula lakes.	Meliadine Lake fish tissue chemistry: **Section 4.7** The Peninsula Lakes study has been removed from Version 3 of the AEMP Design Plan
FEIS GN-1	Agnico Eagle has committed to monitoring water quality during different seasons of the year including under-ice and early spring.	**Section 4.2.3**
Water Licensing Process		
EC-15	Agnico Eagle has committed to providing Benchmarks and Low Action Level management responses	Low Action Levels were updated in the 2018 AEMP (Golder, 2019) Water quality Benchmarks for the AEMP were updated in the 2020 AEMP Report (Azimuth, 2021)
10 KIA-WL-07	Agnico Eagle has committed to collect water quality data (i.e., field water quality profiles and water quality samples) from three stations (in a triangulated arrangement) at approximately 100 m from the diffuser, during the period of discharge.	**Section 4.1.2**
EC-9 and EC-10	Updated the reference area sampling frequencies	Completed in V1 of the AEMP Design Plan. See **Table 4-2** for the frequency of sampling in each area
KIA-WL-16	List of parameters to be analyzed and the minimum acceptable detection limits.	Parameters and detection limits are provided in each of the respective sections of the AEMP Design Plan.
KIA-WL-11	Agnico Eagle has discussed Significance Thresholds and adaptive management in response to reaching an Action Level.	**Section 6.1; Table 6-1**
EC-9 and EC-13	Agnico Eagle has updated the study types for Water Quality Meliadine Lake and Peninsula Lakes programs (i.e., before- after or control impact designs).	Meliadine Lake: **Section 4.2.5** Agnico Eagle recommends moving the Peninsula Lakes study to the Water Quality and Flow Monitoring Plan (Appendix D in the Water Management Plan)
EC-7	Agnico Eagle has provided clarification on the monitoring and adaptive management to be implemented to detect changes and prevent impacts to lake productivity in the effluent mixing area.	Phytoplankton Study: **Section 4.3** Action Levels: **Section 6.2**
EC-12	Clarification on selection of sampling location for fish based on information request from ECCC	**Section 4.6.4** The scope of the fish health studies for the AEMP may be refined based on comments and recommendations received from the Technical Advisory Panel in their review of successive EEM study designs.

Notes:
AEMP = Aquatic Effects Monitoring Program; NIRB = Nunavut Impact Review Board; FEIS = Final Environmental Impact Statement; KIA = Kivalliq Inuit Association; GN = Government of Nunavut; IR = information request.

APPENDIX B
WATER QUALITY SCREENING CRITERIA (MELIADINE LAKE AEMP BENCHMARKS)

Table B-1. Water Quality Screening Values for the Meliadine Lake AEMP (based on data from the 2024 AEMP).

Parameter	Units	DL	Normal Range	FEIS[a]	FWAL[b]	GCDWQ[c]	SSWQO[d]	AEMP Action Level[e]	AEMP Benchmark[f]
Field Measurements									
DO (%)	%	-	-	-	-	-	-	-	-
DO (mg/L)	mg/L	-	-	-	-	-	-	6.5	6.5
pH (field)	pH units	-	7.1 | 7.95	-	6.5 | 9	-	-	6.5 | 9.0	6.5 | 9.0
Sp. Conductivity (field)	uS/cm	-	-	-	-	-	-	-	-
Temperature	C	-	-	-	-	-	-	-	-
Conventional Parameters									
Conductivity (lab)	uS/cm	1	77.5	-	-	-	-	-	-
Hardness	mg/L	0.5	23.4	-	-	-	-	-	-
pH (lab)	pH units	0.1	-	-	6.5 | 9	-	-	6.5 | 9.0	6.5 | 9.0
Total Dissolved Solids	mg/L	10	54	68	500	-	1000	375	500
Total Dissolved Solids (Calculated)	mg/L	1	39.6	68	500	-	1000	375	500
Total Suspended Solids	mg/L	1	1	3.1	-	-	-	-	-
Turbidity (lab)	NTU	0.1	-	-	-	-	-	-	-
Major Ions									
Acidity, Total	mg/L	2	-	-	-	-	-	-	-
Alkalinity, Bicarbonate	mg/L	1	25	-	-	-	-	-	-
Alkalinity, Carbonate	mg/L	1	-	-	-	-	-	-	-
Alkalinity, Hydroxide	mg/L	1	-	-	-	-	-	-	-
Alkalinity, Phenolphthalein	mg/L	1	-	-	-	-	-	-	-
Alkalinity, Total	mg/L	1	20.5	-	-	-	-	-	-
Bromide	mg/L	0.1	-	-	-	-	-	-	-
Calcium (D)	mg/L	0.01	-	-	-	-	-	-	-
Calcium (T)	mg/L	0.01	7.33	-	-	-	-	-	-
Chloride	mg/L	0.1	9.56	14	120	-	-	90	120
Fluoride	mg/L	0.02	0.028	0.0084	0.12	1.5	2.8	2.1	2.8
Magnesium (D)	mg/L	0.004	-	-	-	-	-	-	-
Magnesium (T)	mg/L	0.004	1.18	-	-	-	-	-	-
Potassium (D)	mg/L	0.02	-	-	-	-	-	-	-
Potassium (T)	mg/L	0.02	0.95	-	-	-	-	-	-
Reactive Silica (SiO2)	mg/L	0.01	0.27	-	-	-	-	-	-
Sodium (D)	mg/L	0.02	-	-	-	-	-	-	-
Sodium (T)	mg/L	0.02	4.85	5.3	-	-	-	-	-
Sulphate	mg/L	0.3	3.87	38	128 | 218	-	-	96 | 164	128 | 218
Nutrients									
Ammonia (as N)	mg/L	0.005	0.018	0.54	0.41 | 8.47	-	-	0.308 | 6.35	0.41 | 8.47
Nitrate (as N)	mg/L	0.005	0.018	0.25	2.9	10	-	2.17	2.9
Nitrate + Nitrite (as N)	mg/L	0.0051	-	-	-	-	-	-	-
Nitrite (as N)	mg/L	0.001	0.001	0.051	0.06	1	-	0.045	0.06
Nitrogen	mg/L	0.05	-	-	-	-	-	-	-
Orthophosphate (PO4-P)	mg/L	0.001	0.001	-	-	-	-	-	-
Total Diss Phosphorus	mg/L	0.001	0.00314	-	-	-	-	-	-
Total Dissolved Nitrogen	mg/L	0.05	-	-	-	-	-	-	-
Total Kjeldahl Nitrogen	mg/L	0.05	0.2497	-	-	-	-	-	-
Total Kjeldahl Nitrogen (diss)	mg/L	0.05	-	-	-	-	-	-	-
Total Phosphorus	mg/L	0.001	0.006	0.0049	-	-	-	-	-
Organic/Inorganic Carbon									
Dissolved Organic Carbon	mg/L	0.5	2.72	-	-	-	-	-	-
Total Organic Carbon	mg/L	0.5	3	-	-	-	-	-	-
Total Metals									
Aluminum (T)	ug/L	1	5.32	9.1	271 | 687	-	-	203 | 515	271 | 687
Antimony (T)	ug/L	0.02	0.02	0.51	-	6	-	4.5	6
Arsenic (T)	ug/L	0.02	0.275	3.8	5	10	25	18.8	25
Barium (T)	ug/L	0.02	8.05	77	-	1000	-	750	1000
Beryllium (T)	ug/L	0.005	0.005	-	-	-	-	-	-
Bismuth (T)	ug/L	0.005	0.005	-	-	-	-	-	-
Boron (T)	ug/L	5	6.52	23	1500	5000	-	1120	1500
Cadmium (T)	ug/L	0.005	0.005	0.05	0.04 | 0.065	5	-	0.03 | 0.049	0.04 | 0.065
Cesium (T)	ug/L	0.005	-	-	-	-	-	-	-
Chromium (T)	ug/L	0.1	0.10	1.1	5	50	-	3.75	5
Cobalt (T)	ug/L	0.005	0.016	-	0.78	-	-	0.585	0.78
Copper (T)	ug/L	0.05	0.86	2	-	2000	-	1500	2000
Gallium (T)	ug/L	0.05	-	-	-	-	-	-	-
Iron (T)	ug/L	1	15.0	42	300	-	1060	795	1060
Lanthanum (T)	ug/L	0.01	-	-	-	-	-	-	-
Lead (T)	ug/L	0.01	0.022	0.15	-	5	-	3.75	5
Lithium (T)	ug/L	0.5	0.72	-	-	-	-	-	-
Manganese (T)	ug/L	0.05	3.062	5.5	-	120	-	90	120
Mercury (T)	ug/L	0.5	8.00E-04	0.02	0.026	1	-	0.020	0.026
Molybdenum (T)	ug/L	0.05	0.11	5.2	73	-	-	54.8	73
Nickel (T)	ug/L	0.05	0.44	2.7	25	-	-	18.8	25
Niobium (T)	ug/L	0.1	-	-	-	-	-	-	-
Phosphorus (T)	ug/L	50	-	-	-	-	-	-	-
Rhenium (T)	ug/L	0.005	-	-	-	-	-	-	-
Rubidium (T)	ug/L	0.005	-	-	-	-	-	-	-
Selenium (T)	ug/L	0.04	0.049	0.16	1	50	-	0.75	1

Table B-1. Water Quality Screening Values for the Meliadine Lake AEMP (based on data from the 2024 AEMP).

Parameter	Units	DL	Normal Range	FEIS[a]	FWAL[b]	GCDWQ[c]	SSWQO[d]	AEMP Action Level[e]	AEMP Benchmark[f]
Silicon (T)	ug/L	50	-	-	-	-	-	-	-
Silver (T)	ug/L	0.005	0.005	0.1	0.25	-	-	0.188	0.25
Strontium (T)	ug/L	0.02	36.1	-	2500	7000	-	1880	2500
Sulfur (T)	ug/L	500	-	-	-	-	-	-	-
Tantalum (T)	ug/L	0.1	-	-	-	-	-	-	-
Tellurium (T)	ug/L	0.02	-	-	-	-	-	-	-
Thallium (T)	ug/L	0.005	0.005	0.1	0.8	-	-	0.6	0.8
Thorium (T)	ug/L	0.005	-	-	-	-	-	-	-
Tin (T)	ug/L	0.02	0.038	-	-	-	-	-	-
Titanium (T)	ug/L	0.05	0.17	-	-	-	-	-	-
Tungsten (T)	ug/L	0.01	-	-	-	-	-	-	-
Uranium (T)	ug/L	0.001	0.016	1.5	15	20	-	11.2	15
Vanadium (T)	ug/L	0.05	0.05	-	120	-	-	90	120
Yttrium (T)	ug/L	0.01	-	-	-	-	-	-	-
Zinc (T)	ug/L	0.5	1.70	6.7	-	-	-	-	-
Zirconium (T)	ug/L	0.01	-	-	-	-	-	-	-
Dissolved Metals									
Aluminum (D)	ug/L	1	-	-	-	-	-	-	-
Antimony (D)	ug/L	0.02	-	-	-	-	-	-	-
Arsenic (D)	ug/L	0.02	-	-	-	-	-	-	-
Barium (D)	ug/L	0.02	-	-	-	-	-	-	-
Beryllium (D)	ug/L	0.005	-	-	-	-	-	-	-
Bismuth (D)	ug/L	0.005	-	-	-	-	-	-	-
Boron (D)	ug/L	5	-	-	-	-	-	-	-
Cadmium (D)	ug/L	0.005	-	-	-	-	-	-	-
Cesium (D)	ug/L	0.005	-	-	-	-	-	-	-
Chromium (D)	ug/L	0.1	-	-	-	-	-	-	-
Cobalt (D)	ug/L	0.005	-	-	-	-	-	-	-
Copper (D)	ug/L	0.05	0.861	-	1.61 | 4.92	-	-	1.21 | 3.69	1.61 | 4.92
Gallium (D)	ug/L	0.05	-	-	-	-	-	-	-
Iron (D)	ug/L	1	-	-	-	-	-	-	-
Lanthanum (D)	ug/L	0.01	-	-	-	-	-	-	-
Lead (D)	ug/L	0.01	0.013	-	4.98 | 7.74	-	-	3.73 | 5.8	4.98 | 7.74
Lithium (D)	ug/L	0.5	-	-	-	-	-	-	-
Manganese (D)	ug/L	0.05	1.196	-	210 | 330	-	-	158 | 248	210 | 330
Mercury (D)	ug/L	0.5	-	-	-	-	-	-	-
Molybdenum (D)	ug/L	0.05	-	-	-	-	-	-	-
Nickel (D)	ug/L	0.05	-	-	-	-	-	-	-
Niobium (D)	ug/L	0.1	-	-	-	-	-	-	-
Phosphorus (D)	ug/L	50	-	-	-	-	-	-	-
Rhenium (D)	ug/L	0.005	-	-	-	-	-	-	-
Rubidium (D)	ug/L	0.005	-	-	-	-	-	-	-
Selenium (D)	ug/L	0.04	-	-	-	-	-	-	-
Silicon (D)	ug/L	50	-	-	-	-	-	-	-
Silver (D)	ug/L	0.005	-	-	-	-	-	-	-
Strontium (D)	ug/L	0.02	-	-	2500	-	-	1880	2500
Sulfur (D)	ug/L	500	-	-	-	-	-	-	-
Tantalum (D)	ug/L	0.1	-	-	-	-	-	-	-
Tellurium (D)	ug/L	0.02	-	-	-	-	-	-	-
Thallium (D)	ug/L	0.005	-	-	-	-	-	-	-
Thorium (D)	ug/L	0.005	-	-	-	-	-	-	-
Tin (D)	ug/L	0.02	-	-	-	-	-	-	-
Titanium (D)	ug/L	0.05	-	-	-	-	-	-	-
Tungsten (D)	ug/L	0.01	-	-	-	-	-	-	-
Uranium (D)	ug/L	0.001	-	-	-	-	-	-	-
Vanadium (D)	ug/L	0.05	-	-	-	-	-	-	-
Yttrium (D)	ug/L	0.01	-	-	-	-	-	-	-
Zinc (D)	ug/L	0.5	1.90	-	6.87 | 14.2	-	-	5.16 | 10.6	6.87 | 14.2
Zirconium (D)	ug/L	0.01	-	-	-	-	-	-	-
Cyanides									
Cyanide (Free)	mg/L	0.001	-	0.00035	-	-	-	-	-
Cyanide (Total)	mg/L	0.001	0.001	0.009	0.005	0.2	-	0.00375	0.005
Cyanide (WAD)	mg/L	0.001	-	-	-	-	-	-	-

Notes:
[a] FEIS predictions for the edge of the mixing zone as presented in Agnico Eagle (2014).
[b] The freshwater aquatic life guidelines (FWAL) for aluminum (T), cadmium (T), copper (D), lead (D), manganese (D), and zinc (D) are variable depending on modifying factors such as pH, hardness, and DOC. Values shown represent the range of FWAL guidelines calculated for MEL-01 open-water samples in 2024.
[c] Guidelines for Canadian Drinking Water Quality - Health Canada drinking water guidelines (maximum acceptable concentrations).
[d] Site-specific water quality objectives for fluoride, arsenic, and iron.
[e] The AEMP Action Level is 75% of the AEMP Benchmark.
[f] The AEMP Benchmark is the lowest of the FWAL or GCDWQ.

APPENDIX C
RESPONSE TO COMMENTS FROM THE AGENCIES ON VERSION 2 OF THE AEMP DESIGN PLAN (JANUARY 2023)

This document presents responses to comments that were received from Environment and Climate Change Canada (ECCC) on the AEMP Design Plan (Draft for Discussion) that was submitted to the Nunavut Water Board with in the 2021 Annual Report. Comments on the AEMP Design Plan and the 2021 AEMP Report were provided to Azimuth Consulting Group Inc (Azimuth) in an email from the Meliadine Environment Department on July 3, 2022. Azimuth provided written responses by email to the Meliadine Environment Department on July 12, 2022. The comments and response specific to the AEMP Design Plan are provided below.

ECCC-3 Definitions for IC25 and QA/QC Blanks

Reference(s)

- Appendix 32-1 AEMP Design Plan
- List of Abbreviations
- Section 5.1.5 Quality Assurance/Quality Control

Comment

IC25 – The ICp is the inhibiting concentration for a specified percent effect, such as a 25% reduction in growth. The definition for IC25 provided should be corrected from “inhibition concentration affecting 25% of tested organisms” to “effluent concentration that causes a 25% inhibitory effect in the sublethal endpoint being measured”. The definition provided is for EC25 rather than IC25.

QA/QC – Errata note: The descriptions of travel and field blanks in the AEMP Design QA/QC section on page 44 have been transposed and should be corrected.

ECCC Recommendations(s)

ECCC recommends revising the definitions as noted, for clarity.

Response

The definition of the IC25 has been updated as requested.

The descriptions of travel and field blanks were corrected.

ECCC-5 Low Action Levels – Phytoplankton Assessment Criteria

Reference(s)

- Appendix 32-1 AEMP Design – Table 8-2 Proposed Low Action Levels for Toxicological Impairment for Meliadine Lake

Comment

The first part of the Phytoplankton Assessment Criteria is “Phytoplankton community metrics at the Near-field area beyond the range of baseline/reference conditions”

For toxicological impairment, most of the metrics would demonstrate a lower value (e.g. density and biomass), but using the descriptive term “beyond” implies higher. This should be clarified by describing the trigger as “below” or “outside” the range of baseline/reference conditions.

Footnote (c) is missing for this table.

ECCC Recommendations(s)

ECCC recommends revision of the assessment criteria statement to specify “below” or “outside” rather than “beyond” the range of baseline/reference conditions and that footnote (c) be completed.

Response

We agree with ECCC’s recommendation. We have revised the assessment criteria to state “outside the range of baseline/reference conditions”.

ECCC-6 Proposed Action Levels for Nutrient Enrichment Hypothesis

Reference(s)

- Appendix 32-1 AEMP Design – Table 8-3 Proposed Action Low Action Levels for Nutrient Enrichment for Meliadine Lake

Comment

In order to meet the Low Action Level for Water Quality, the following three conditions are proposed to have to exist:

- Concentrations of TP in the Near-field area above the normal range, supported by temporal trends AND
- A statistically significant relative difference between the Near-field area and Reference for TP AND
- Lake-wide average phosphorus concentration exceeds 75% of AEMP Benchmark

Considering the extent and volume of Meliadine Lake, the third condition would almost certainly never be measured, and to be met would entail an increase of significant magnitude in TP loadings and ensuing concentrations. The AEMP Benchmark has been set at 0.010 mg/L TP to reflect the upper bound of the oligotrophic status, and the Action level trigger would be 0.0075 mg/L TP. A more timely and realistic trigger condition would be on the basis of near-field rather than lake-wide change.

ECCC Recommendations(s)

ECCC recommends amending the third condition by replacing “lake-wide” with “near-field”.

Response

The AEMP Action Level for phosphorus will be applied to the near-field area. However, we want to emphasize that phosphorus concentrations are one of the lines of evidence used to assess nutrient enrichment caused by effluent. Increases in total phosphorus in the East Basin suggests the potential for nutrient enrichment, but any conclusions about the potential for nutrient enrichment need to be supported by more relevant lines of evidence that directly assess phytoplankton productivity, namely total biomass and chlorophyll-a concentrations.

APPENDIX D
RESPONSE TO COMMENTS FROM THE AGENCIES ON VERSION 3 OF THE AEMP DESIGN PLAN (JANUARY 2024)

This document presents responses to comments that were received from Environment and Climate Change Canada (ECCC) on Version 3 of the AEMP Design Plan that was submitted to the Nunavut Water Board in January 2024. Responses from the agencies were received on May 10, 2024.

ECCC-TC-08 Water quality screening criteria for parameters without CCME guidelines

Request Made by Interested Party:

ECCC recommends the Proponent update the water quality screening criteria in both the Water Balance and Water Quality Model and Aquatic Effects Monitoring Plan Design Plan, to include Federal Environmental Quality Guidelines for cobalt, copper, strontium and vanadium.

Agnico Eagle's Response to Request:

Agnico Eagle will update the Aquatic Effects Monitoring Plan Design Plan to include Federal Environmental Quality Guidelines for cobalt, copper, strontium, and vanadium. The update plan will be provided 60 days after issuance of the Amended Water Licence.

ECCC-TC-13 Aquatic effects monitoring program monitoring peninsula lakes

Request Made by Interested Party:

ECCC recommends the Proponent retain monitoring of the D7 peninsula lake in the Aquatic Effects Monitoring Program Design Plan and propose alternative lakes for monitoring when lakes A8 and B7 will be dewatered, so that a robust monitoring program continues for the peninsula lakes.

Agnico Eagle's Response to Request:

Instead of moving Lake D7 to the Water Quality and Flow Monitoring Plan, Agnico Eagle will keep Lake D7 in the AEMP. ECCC recommended Agnico Eagle add two new lakes to the AEMP to replace Lake A8 and Lake B7. We agree that additional lakes should replace A8 and B7. Instead of adding new lakes to the study design, we recommend leveraging the existing compliance monitoring dataset for Lake E3 (MEL-15), Lake G2 (MEL-16), and Lake H1 (MEL-17). Other than A8, B7, and D7, no other lakes on the peninsula are monitored more frequently than E3, G2, and H1.

The updated AEMP will be provided 60 days after issuance of the Amended Water Licence which would include the two new peninsula lakes from Lake E3 (MEL-15), Lake G2 (MEL-16), or Lake H1 (MEL-17), plus Lake D7.

ECCC-TC-14 Benthic community measurement endpoint

Request Made by Interested Party:

ECCC recommends the Proponent justify why they will no longer be using "Benthic community similarity between exposure and reference areas" as a measurement endpoint in the Aquatic Effects Monitoring Program.

Agnico Eagle's Response to Request:

Between Version 2 (Table 5-7) and Version 3 (Table 4.10) benthic community similarity is included and the reader should refer to these tables.

Measurement endpoints for the benthic invertebrate community study are provided in Table 4-10 Aquatics Effects Monitoring Program (AEMP) Design Plan Version 3; this table states that Bray-Curtis is an "AEMP Variable" for the benthic invertebrate community study.

Table 3-1 in the AEMP Design Plan is intentionally generic. The table is meant to highlight how the conceptual model and problem formulation stages helped design the AEMP. The table is not meant to provide a comprehensive and detailed accounting of the various endpoints and statistical methods used for each component of the AEMP. The reader should refer to Table 4.10 (Version 3) for the specific benthic analysis and as stated in Section 4.5.1 the objectives of which includes similarities between expose and reference areas.

ECCC-TC-16 Stickleback study

Request Made by Interested Party:

ECCC recommends the Proponent clarify if lethal threespine stickleback population studies will be done for the Aquatic Effects Monitoring Program, and if not, then justify why the proposed AEMP fish population study differs from that proposed for the Environmental Effects Monitoring Program.

Agnico Eagle's Response to Request:

Agnico Eagle initially considered a non-lethal study for the Threespine Stickleback program, but ultimately decided that a lethal study was the most scientifically defensible option to assess the health of this population. The AEMP Design Plan (January 2024 submitted with the Application) was submitted before we finalized the Cycle 3 EEM study design (February 2024), hence the discrepancy between the two documents. The AEMP Design Plan does mention that the Threespine Stickleback study may be revised pending review of the Cycle 3 EEM Technical Advisory Panel. We hope to hear from the Technical Advisory Panel before the end of May.

ECCC-TC-17 Parameter concentration normal ranges in Meliadine Lake

Request Made by Interested Party:

ECCC recommends the Proponent explain:

a. the rationale or explanation for changing the dates/periods of data for calculating normal water quality ranges,

b. why different dates/periods are now used for the reference and other areas, and

c. how these new data dates/periods change the calculated normal.

Agnico Eagle's Response to Request:

Responses a) b), and c)

The Normal Ranges for Meliadine Lake were updated in the 2020 AEMP to include reference area samples from MEL-03, MEL-04, and MEL-05 in 2019 and 2020. In addition, the Normal Ranges that were calculated in 2018 were provisional, and authors expressly stated that the Normal Ranges would be updated to include new reference area data (see page iii in the Executive Summary of the 2018 AEMP and Cycle 1 EEM [Golder 2019]).

The refined normal ranges described in the AEMP Design Plan (Section 4.3.4) have not changed since 2020 and have been used in the previous AEMP annual reports (including the 2020, 2021, 2022, and 2023 AEMP annual report). Therefore, Agnico Eagle feels this is an approved methodology and an approved set of normal range values.

ECCC-TC-18 Comparison between observations and FEIS predictions

Request Made by Interested Party:

ECCC recommends the Proponent update the Aquatic Effects Monitoring Program Design Plan, with a continuation of comparing observed water quality at MEL-1 against the FEIS predictions, and the addition of a comparison of observed water quality at MEL-1 against updated models.

Agnico Eagle's Response to Request:

Agnico Eagle has done this comparison in the past and most recently in the 2023 Annual Report and will continue to do so in the future. Based on this, the AEMP Design Plan will be updated to reflect this and will be submitted 60 days after issuance of the Amended Water Licence.